REVIEW



Cancer-nervous system crosstalk: from biological mechanism to therapeutic opportunities



Sirui Huang^{1†}, Jing Zhu^{1†}, Linglu Yu¹, Yan Huang^{2*} and Yue Hu^{1,3,4*}

Abstract

A growing body of research suggests a bidirectional interaction between cancer and the nervous system. Neural cells exert their effects on tumors by secreting neurotransmitters and cell adhesion molecules, which interact with specific receptors on tumor cells to modulate their behavior. Conversely, tumor-secreted factors, particularly including inflammatory factors, can alter neural activity and increase neuronal excitability, potentially contributing to neurological manifestations such as epilepsy. The immune system also serves as a crucial intermediary in the indirect communication between cancer and the nervous system. These insights have opened promising avenues for novel therapeutic strategies targeting both tumors and their associated neurological complications. In this review, we have synthesized the key biological mechanisms underlying cancer-nervous system interactions that have emerged over the past decade. We outline the molecular and cellular pathways mediating this cross-talk and explore the clinical implications of targeting the nervous system to suppress tumor growth and metastasis, mitigate neurological complications arising from cancer progression, and modulate the immune response through neural regulation in the context of cancer therapy.

Keywords Cancer, Neurobiology of cancer, Oncological therapeutic modalities, Nervous system

[†]Sirui Huang and Jing Zhu contributed equally to this work

*Correspondence: Yan Huang jacob6666@163.com Yue Hu yuehu@njucm.edu.cn Full list of author information is available at the end of the article



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Background

The recognition that the nervous system plays an active and complex role in tumor growth and progression represents a pivotal advancement in the field of oncology [1]. Early observations of the interaction between the nervous system and tumors date back to the latter half of the nineteenth century. During this period, autopsies of patients with advanced esophageal and tongue cancers revealed that tumor expansion could compress and damage adjacent nerves, resulting in neurological dysfunction [2, 3]. At that time, nervous tissue was largely regarded as a passive structure, much like muscle or connective tissue, that was secondarily affected by tumor invasion. Consequently, the potential active role of the nervous system in cancer progression remained largely underappreciated [4-7]. In the ensuing decades, researchers gradually identified phenomena such as perineural invasion (PNI) [8, 9] and demonstrated that tumor-derived factors could induce peripheral nerve inflammation even in regions not directly invaded by the tumor [10]. Concurrently, studies revealed that neurotrophic factors secreted by nerves, such as nerve growth factor (NGF) and brain-derived neurotrophic factor (BDNF), could facilitate tumor proliferation and survival [11]. However, despite these findings, the conceptual framework for understanding neurotumoral interactions remained relatively stagnant. The real turning point came in 2013, when Magnon et al. demonstrated for the first time in a mouse model of prostate cancer that nerve fibers can grow out of tumor tissue through the process of axonogenesis, thereby promoting cancer growth and spread [12]. Subsequent validation in mouse models of breast [13], gastric [14], and lung cancers [15] has confirmed the presence of infiltrating autonomic and sensory nerve fibers within tumor tissues, releasing neurotransmitters that engage with their specific receptors on both stromal and cancer cells [16, 17]. These discoveries have transformed our understanding of cancer-nerve interactions and have given rise to a new interdisciplinary field: cancer neurobiology.

The emerging field of cancer neurobiology has garnered increasing attention in recent years, as the pivotal role of the nervous system in cancer development and progression has become increasingly evident. Recent research indicates that neural activity is crucial for the development of hallmark cancer capabilities [18]. Specifically, neuronal activity has been shown to modulate the immune response and facilitate cancer cells' ability to escape immune surveillance [19]. Moreover, accumulating evidence has uncovered multiple modes of communication between cancer cells and neurons, highlighting the complexity of their bidirectional interactions. Particular attention has also been directed toward the interplay between the nervous system and the tumor microenvironment, which plays a pivotal role in shaping tumor behavior and therapeutic response [18, 20]. This evolving understanding suggests that targeting neuraltumor interactions represent a promising therapeutic strategy. For instance, denervation approaches have been shown to slow or even halt tumor growth in preclinical models. Additionally, pharmacological inhibition of specific neurotransmitter receptors, such as M3 muscarinic and metabotropic glutamate receptor 1 (mGluR1), has demonstrated efficacy in suppressing tumor progression, thereby emerging as a viable oncological intervention [21]. Advancements in cancer neurobiology are thus essential for elucidating the intricate crosstalk between the nervous system and cancer. They provide novel insights into tumor biology and pave the way for innovative treatment strategies. The objective of this review is to elucidate the molecular mechanisms underlying neuraltumor interactions, outline the principal signaling pathways involved, and summarize recent breakthroughs in the field. Furthermore, the review evaluates the therapeutic potential of newly developed neural-targeted interventions and explores their implications for improving clinical outcomes in cancer patients.

Interactions between cancer and the nervous system

The role of neural activity in cancer progression Neuronal activity promotes cancer growth

Neuronal activity contributes to tumor development and growth through various mechanisms, such as direct interaction with tumor cells, indirect modulation of the immune system and alterations in tumor vascularization. Current research predominantly focuses on the roles of neurotransmitters and neurotrophic factors secreted by neurons, which bind to specific receptors on tumor cells and directly influence their growth and survival. Neurotransmitters in the synaptic cleft, such as adrenaline, norepinephrine, dopamine, acetylcholine, glutamate, etc., along with neurotrophic factors, such as BDNF, NGF, Neurotrophin-3, etc., and neurogenic chemokines, such as C-X-C motif chemokine ligand 12 (CXCL12) and C-X3-C motif chemokine ligand 1 (CX3CL1), etc., can interact with corresponding receptors on tumor cells (Fig. 1A). These interactions activate a range of downstream signaling cascades that support tumor growth, invasion, and metastasis [22, 23]. Tumor cells frequently exhibit dysregulated expression and function of ion channels and neurotransmitter receptors, rendering them more susceptible to modulation by neuronal signals. This dysregulation contributes to unchecked proliferation and dissemination [24, 25]. For example, the excitatory neurotransmitter glutamate activates the N-methyl-Daspartate receptor (NMDAR) on tumor cells, triggering downstream pathways such as Calcium/Calmodulin-Dependent Protein Kinase II/IV (CaMKII/IV) and mitogen-activated extracellular signal-regulated kinase (MEK)-mitogen-activated protein kinase (MAPK). These pathways ultimately lead to the activation of the transcription factor cAMP response element-binding protein (CREB), promoting tumor growth and invasiveness [26] (Fig. 1B). Similarly, neurotrophic factors, including BDNF, NGF and Neurotrophin-3, activate neurotrophic factor receptors like p75 neurotrophin receptor (p75 NTR), tropomyosin receptor kinase B (TrkB), and tropomyosin receptor kinase C (TrkC) on tumor-initiating cells, enhancing their vitality through the extracellular signal-regulated kinase (ERK) and Ak strain transforming (AKT) pathways [27, 28] (Fig. 1B).

Building on the direct effects of neuronal activity on tumor cells, it is also important to explore the indirect

⁽See figure on next page.)

Fig. 1 Different pathways of neural influence on tumors: **A**. Neurons act on tumor cells via neurotransmitters, neurotrophic factors, neurogenic chemokines, etc., through corresponding receptors. **B**. By secreting glutamate, neurons bind to glutamate receptors, such as NMDAR, AMPAR, etc., on tumors to promote tumor growth and invasion. Neurons can also promote tumor growth and invasion by secreting neurotrophic factors that bind to tumor receptors. **C**. Neurons secrete neurotransmitters such as acetylcholine and GABA to act on tumor cells, triggering tumor immune escape. **D**. Neurons secrete MDK to activate CD8⁺T cells and promote the secretion of CCL4, enhancing CCL5 expression in microglial cells, thereby mediating cancer cell proliferation. **E**. Neurons promote angiogenesis in the tumor microenvironment by secreting norepinephrine (NE), providing nutrients for tumor growth. **F**. Tumor cells form a network through TM and TNT, where neurons may regulate TM and TNT formation. **G-I**. The interaction mechanism between the nervous system and tumors. **G**. Schwann cells envelop cancer cells, forming synchronized tissue patterns of Schwann cells and cancer cells, termed TAST. **H**. Activated Schwann cells secrete CL2, recruiting monocytes that differentiate into macrophages and secrete cathepsin B, promoting PNI of tumors. **I**. Schwann cells secrete IL-6 and CXCL5, acting on tumor cells to activate STAT3 and Phosphatidylinositol 3-kinase (PI3K)/AKT/GSK-3β pathways, enhancing cell migration



Fig. 1 (See legend on previous page.)

roles neurons play in modulating the tumor microenvironment, particularly in facilitating immune evasion. During tumor growth, cancer cells acquire the ability to evade immune surveillance, thereby avoiding recognition and elimination by the host immune system. Neurons contribute to this process not only by acting directly on tumor cells to induce immune escape mechanisms but also by modulating the function of immune cells within the tumor microenvironment. This neuronal influence diminishes the immune system's capacity to effectively detect and eliminate malignant cells. For example, the neurotransmitter acetylcholine has been shown to upregulate the expression of the immune checkpoint molecule Programmed cell death 1 ligand 1 (PD-L1) on tumor cells, thereby conferring resistance to cytotoxic CD8⁺ T cells [29] (Fig. 1C). Similarly, the inhibitory neurotransmitter γ -Aminobutyric Acid (GABA) activates GABA_B receptors on cancer cells, suppressing Glycogen Synthase Kinase 3β (GSK-3 β) activity, thereby enhancing β -catenin signaling. This cascade not only promotes tumor cell proliferation but also impedes CD8⁺ T cell infiltration into the tumor microenvironment [30] (Fig. 1C). Moreover, neurons can produce Midkine (MDK), a neurotrophic factor that modulates T cell activity. MDK stimulation leads to increased expression of the pro-inflammatory chemokine C-C motif chemokine ligand 4 (CCL4) by CD8⁺ T cells. This activation of CD8⁺ T cells and subsequent production of CCL4 result in insufficient T cells in the tumor microenvironment to recognize and eliminate tumor cells. Additionally, the production of CCL4 can promote the nuclear factor KB (NF-KB)dependent expression of C-C motif chemokine ligand 5 (CCL5) by microglial cells, which in turn activates the AKT/GSK-3β/CREB pathway, promoting tumor cell proliferation and resistance to apoptosis [18, 31] (Fig. 1D). In addition to immune modulation, neuronal activity also supports tumor growth by promoting angiogenesis, ensuring a continuous nutrient supply. Sympathetic nerve-derived norepinephrine activates adrenoceptor ß2 (ADRB2) on endothelial cells, reprogramming their metabolism toward aerobic glycolysis to facilitate angiogenesis within the tumor microenvironment [32, 33] (Fig. 1E).

In summary, neuronal activity regulates both direct and indirect regulatory effects on tumor biology, influencing cancer progression through multiple interrelated mechanisms. Although cancer neurobiology is still in its infancy, growing evidence has unveiled diverse neural pathways that contribute to tumor development. These insights offer promising avenues for therapeutic intervention. Future research should focus on elucidating the specific roles of various neuronal and glial subtypes in tumor-nervous system interactions. Moreover, understanding the differential contributions of the central versus peripheral nervous systems to cancer progression represents a critical direction for further investigation.

Neuronal activity promotes cancer metastasis

In addition to its role in tumor initiation and proliferation, neuronal activity significantly contributes to cancer metastasis by enhancing tumor cell invasiveness. Neurons secrete a variety of neurotransmitters and neurotrophic factors that modulate gene expression and facilitate direct communication between tumor cells, thereby increasing malignancy and metastatic potential. For example, norepinephrine can upregulate human telomerase reverse transcriptase (hTERT) expression via the ADRB2/p21-activated kinases (PAK)/Src/hypoxiainducible factor 1α (HIF- 1α) and cellular MYC (c-MYC) signaling pathways, promoting epithelial-mesenchymal transition (EMT) and facilitates the invasion of ovarian cancer cells.

Beyond molecular signaling, tumor cells can form specialized membrane-bound tubular structures: such as tumor microtubes (TMs) and tunneling nanotubes (TNTs), which allow long-distance intercellular communication and contribute to cancer invasion, therapy resistance, and metastasis [33, 34] (Fig. 1F). Under stress conditions, tumor cells generate these structures to transfer cellular components, including organelles, RNA, proteins, and signaling molecules, to neighboring or damaged cells, thereby enhancing the resilience and survival of recipient cells [35]. In gliomas, the formation of TMs has been linked to neuron-associated proteins such as growth-associated protein 43 (GAP-43) and tweety-homolog 1 (Ttyh1), which also facilitate microtubule-dependent tumor cell proliferation, invasion, interconnectivity, and radioresistance [36]. Although a direct regulatory role of neurons in TM or TNT formation has yet to be confirmed, the involvement of neuron-associated proteins suggests potential crosstalk between neural signaling and these metastatic pathways. Targeting TM and TNT formation mechanisms, especially those involving neural components, could complement traditional cancer therapies by reducing metastatic spread and recurrence.

Non-neuronal cells of the nervous system, particularly glial cells, also play essential roles in supporting tumor metastasis. Schwann cells, a key component of the peripheral nervous system, undergo morphological and metabolic reprogramming that facilitates tumor migration and invasion [37, 38]. Within the pancreatic cancer microenvironment, Schwann cells are reprogrammed via c-Jun-dependent pathways typically associated with nerve injury, transitioning to a non-myelinating phenotype. These reprogrammed Schwann cells enwrap pancreatic cancer cells, forming organized cellular columns that guide tumor cells into aligned chains, thereby establishing migratory routes known as tumor-activated Schwann cell trajectories (TAST) [37] (Fig. 1G). In addition to providing structural guidance, these demyelinated Schwann cells can exert mechanical tension to further accelerate tumor dissemination. Activated Schwann cells secrete the chemokine C-C motif chemokine ligand 2 (CCL2), which recruits inflammatory monocytes that differentiate into macrophages capable of expressing the extracellular protease cathepsin B. These macrophages, along with cathepsin B, contribute functionally to PNI, along with cathepsin B, contribute functionally (Fig. 1H). Moreover, co-culture studies have demonstrated that Schwann cell-derived interleukin-6 (IL-6) promotes pancreatic cancer cell migration and invasiveness via activation of the Signal Transducer and Activator of Transcription 3 (STAT3) signaling pathway. This effect is abrogated by IL-6 neutralization or STAT3 downregulation, confirming the functional role of this axis in tumor progression [39] (Fig. 1I). Similarly, Schwann cells can secrete CXCL5, which interacts with CXCR2 receptors on lung cancer cells. This interaction activates the PI3K/ AKT/GSK-3β pathway, upregulating EMT regulators such as Snail and Twist and thereby enhancing cancer cell motility and metastatic potential [40].

Collectively, these findings underscore the multifaceted role of the nervous system in facilitating cancer metastasis. Both neuronal and glial components actively participate in shaping a pro-metastatic microenvironment through molecular signaling, structural remodeling, and immune modulation. Elucidating the precise mechanisms of neural regulation in cancer dissemination may pave the way for novel therapeutic strategies aimed at disrupting these neural-tumor interactions. More detailed information about neuronal activity in cancer progression and metastasis is provided in the table (Table 1).

Cancer impact on neural function

Cancer-released neurotrophins induce nerve outgrowth

Tumors can significantly influence the nervous system by releasing neurotrophins and axon guidance molecules that promote nerve growth and axonogenesis [11, 20]. These factors contribute to the sprouting of new nerve fibers within malignant tissues, effectively linking the tumor microenvironment to the nervous system [17]. NGF, a classical neurotrophin essential for neuronal growth and development, is aberrantly expressed in various types of tumor cells and serves as a biomarker of tumor progression [60]. In a mouse model of pancreatic ductal adenocarcinoma (PDAC), elevated expression and secretion of both NGF and BDNF have been observed [61]. Similarly, increased NGF levels have been reported in colorectal, breast and pancreatic cancers [62, 63], and these tumors exhibit a markedly higher density of newly formed sympathetic nerve fibers compared to their normal tissue counterparts [64]. As tumors expand, these newly formed nerves can extend toward and integrate with the central nervous system. BDNF, another critical neurotrophic factor, regulates synaptic plasticity and promotes the growth, differentiation, and maturation of both central and peripheral neurons [65, 66]. High expression of BDNF in tumor tissues has been correlated with increased tumor invasiveness and a higher propensity for brain metastasis [67]. In pancreatic cancer, tumor-derived BDNF activates Trk receptors, stimulating neuronal axonogenesis and altering neural composition within the brain. Collectively, these findings underscore the role of neurotrophins such as NGF and BDNF as molecular bridges between tumors and the nervous system, facilitating aberrant neural remodeling (Fig. 2A).

In addition to neurotrophins, axon guidance molecules play a pivotal role in tumor-nerve crosstalk. These molecules, initially characterized for their role in directing neuronal axon outgrowth, are now known to be closely associated with tumor development and progression [68]. Key axon guidance families include Plexins/Semaphorins, Erythropoietin-producing hepatocellular carcinoma (Eph)/Eph-family receptor-interacting proteins (Ephrin), Netrins and their receptors, Slit-Robo, all of which are expressed in various malignancies and contribute to cancer cell invasion, migration, and metastasis [69, 70]. For example, in prostate cancer, tumor cells were found to express high levels of semaphorin 4F (S4F), an axon guidance molecule that induces neurite sprouting and elongation when co-cultured with neurons, indicating its role in promoting neural innervation of tumors [71] (Fig. 2B). Another prominent axon guidance system, the Eph/Ephrin axis, comprises Eph receptor tyrosine kinases and their membrane-bound ephrin ligands. This system is highly expressed in tumor tissues and is linked to angiogenesis, invasion, metastasis, and the maintenance of cancer stem cell populations. Recent studies have demonstrated that tumor-derived exosomes enriched with ephrin-B1 can stimulate neurite outgrowth in PC12 cells, suggesting that Eph/Ephrin signaling also contributes to tumor innervation [72] (Fig. 2B). Similarly, netrin-1, a molecule implicated in axonal pathfinding, is highly expressed in many aggressive cancers and binds to the deleted in colorectal cancer (DCC) receptors to activate axon regeneration pathways [73]. In the peripheral nervous system, Schwann cells can utilize netrin-1 as a guidance cue via DCC receptors expressed on regenerating axons, facilitating neural repair and potentially supporting PNI in cancer [74] (Fig. 2C).

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System	Cancer Type	Subtype/Classification	Neuronal Activity Mechanism	Related Molecules/Pathways
Digestive System	Gastric Cancer	Adenocarcinoma	CGRP peptidergic neurons directly connect to gastric cancer spheroids. Chemogenetic activation of these neurons induces calcium release into the cytoplasm of cancer cells, thereby promoting tumor growth and metastasis [41]	CGRP, Ca ²⁺
	Colorectal Cancer		Overexpression of NGF during colorectal carcino- genesis activates the TrkA-MAPK/ERK pathway, upregulates NGAL expression, enhances MMP2 and MMP9 activity, and promotes colorectal cancer metastasis [42]	NGF, Trka-MaPK/ERK, NGAL, MMP2, MMP9
	Liver Cancer	Intrahepatic cholangiocarcinoma (ICC)	Parasympathetic-derived acetylcholine induces epithelial-mesenchymal transition (EMT), thereby promoting ICC metastasis [43]	CAMKII, GSK3β/β-catenin, BDNF
	Pancreatic Cancer	Ductal Adenocarcinoma (PDAC)	Neurogenic chemokines CXCL12 and CX3CL1 facili- tate tumor cell invasion of nerves via their respective receptors CXCR4 and CX3CR1 [44, 45]	CXCL12, CX3CL1, CXCR4, CX3CR1
			The chemokine CCL7, produced by Schwann cells, enhances PDAC cell migration, invasion, and rissue	CCL7, CCR1/STAT2, CD63/PI3K/AKT

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ΛF	CR1

NE, ADRB2/PKA/STAT3, NGF, MMP2/9 ther stimulates Schwann cell proliferation and migraexpression via the CCR1/STAT2 pathway. TIMP1 furinhibitor of matrix metalloproteinases 1 (TIMP1) promoting the invasion and metastasis of cancer Catecholamines, such as NE, released from sympathetic nerves act on ADRB2 in PDAC cells, tion through CD63/PI3K/AKT signaling [46]

through the activation of the ADRB2/PKA/STAT3 signaling pathway. This process also increases the production of NGF and MMP2/9 [47]

axis; disruption of this pathway promotes tumor cell Neuronal activity supports SCLC cell survival by activating the $\mbox{Ga}_{s}/\mbox{cAMP}/\mbox{PKA}/\mbox{CREB signaling}$ death via JNK/c-Jun activation [48]

Small Cell Lung Cancer (SCLC)

Respiratory System Lung Cancer

Ga_s/cAMP/PKA/CREB, JNK/c-Jun

DLL3, KEAP1/NRF2/Notch DLL3 in SCLC does not improve survival, suggest-ing that tumor-associated neuronal markers may not always be effective therapeutic targets [49, 50] Targeting the neuronally expressed Notch ligand Non-Small Cell Lung Cancer (Adenocarcinoma)

NF-kB pathway Neuronatin (NNAT) promotes NSCLC progression Lung cancer cells may promote immune evasion by enhancing tumor innervation via NGF [51]

by inducing NK cell apoptosis through the neuronal-like p75 NTR-proNGF-sortilin signaling axis [52, 53]

proNGF/p75 NTR

Table 1 (continu	(pər			
System	Cancer Type	Subtype/Classification	Neuronal Activity Mechanism	Related Molecules/Pathways
Urinary System	Prostate Cancer	Neuroendocrine Type	The upregulated NGF promotes NEPC development by integrating CHRM4 and activating the AKT-MYCN pathway [54]	NGF-CHRM4 axis
			In the prostate cancer microenvironment, the loss of P-adrenergic signaling exerts a significant inhibi- tory effect on both tumor growth and angiogenesis [32]	Adrenergic nerves, ADRB2 signaling, Coa6
			Autonomic Nerve Development effects the Progression of Prostate Cancer [12]	Adrenergic signals, ADRB2 and ADRB3, Cholinergic signals, CHRM1
	Renal Cancer		Neuron-derived neurotrophic factor suppresses RCC progression by inhibiting EMT, reducing cell proliferation, and promoting G1 to S phase cell cycle arrest [55]	Cyclin D1, Vimentin, MMP-7, Wnt/β-catenin signaling, Ras signaling
Endocrine System	Breast Cancer	Triple-Negative Breast Cancer (TNBC)	Neurons bind to β 2-AR on TNBC by secreting NE, promoting the progression of cancer [56]	NE/β2-AR signaling
			Neuronal proNGF promotes brain metastasis of tri- ple-negative breast cancer cells through the TrkA/ EphA2 pathway [57]	Trka/EphA2
			Parathympathetic nerve secretes BDNF and NGF, acts on TrkA and TrkB receptors on breast cancer cells, activates downstream Akt/PI3K, Jak/STAT, NF-kB, Wht/β-catenin pathways, and promotes cancer migration, proliferation, and angiogenesis [58]	Akt/PI3K, Jak/STAT, NF-kB, Wnt/β-catenin pathways
			GABA secreted by neurons binds to GABAR on breast cancer, activates downstream ERK1/2 pathway, and promotes cancer metastasis [59]	ERK1/2 pathway



Fig. 2 The mechanisms of cancer cell impact on neural function: A. Tumor cells release neurotrophic factors (such as NGF and BDNF) that bind to receptors on neurons, promoting axonogenesis and nerve growth. B. Tumor cells secrete axon growth factors, such as S4F and Ephrin-B1, which bind to corresponding receptors on neurons, facilitating axonogenesis. C. Tumor-derived Netrin-1 binds to DCC receptors expressed on neuronal axons and Schwann cells, guiding axonal regeneration across peripheral nerve bridges

Beyond secreted factors, tumors may promote nerve development through alternative mechanisms. In small cell lung cancer, axonogenesis-related genes and neural migration pathways are upregulated in malignant epithelial cells. These cancer cells exhibit morphological changes reminiscent of neuronal cells and form nervelike intercellular connections. Furthermore, a subset of cancer stem cells (CSCs) isolated from gastric and colorectal cancer patients has been shown to differentiate into neuron-like cells in vitro, including both sympathetic and parasympathetic subtypes, indicating that tumors may actively participate in neurogenesis [75]. Therefore, future research into cancer-neuron interactions may entail examining the morphological and functional analogies between cancer cells and neurons, along with the identification of their principal pathways.

Cancer-derived neuropeptides cause neuropathic pain

Pain is one of the most prevalent and debilitating symptoms experienced by cancer patients, substantially compromising their quality of life. A substantial body of research now substantiates the inextricable link between cancer-induced pain and the infiltration of macrophages [76–78]. Among the mediators of this pain, cancerderived neuropeptides such as calcitonin gene-related peptide (CGRP) and neuropeptide Y (NPY), have been shown to hyperactivate injured peripheral nerve endings, thereby triggering neuropathic pain [79–81]. Moreover, tumors generate excessive glycolytic metabolites, creating a highly acidic tumor microenvironment. This acidification influences the function of cation channels on peripheral sensory neurons, leading to their sensitization and the development of cancer-induced bone pain [82]. In the following sections, we elaborate on the molecular mechanisms underlying tumor-nerve crosstalk and its role in cancer-associated pain.

Neurotrophic factor plays a pivotal role in the pathogenesis of cancer pain. In pancreatic cancer, both malignant and infiltrating immune cells release NGF, which binds to TrkA on sensory neurons, promoting neuronal activation and recruitment [83] (Fig. 3A). This process facilitates perineural infiltration, thereby inducing pain. In the Methyl-N-nitrosourea-induced Rat Mammary Tumor-1 (MRMT-1) model of bone cancer, NPY expression is significantly upregulated, further supporting the involvement of neuropeptides in tumor-induced pain [84]. Furthermore, NGF, along with other neurotrophic factors, can activate the transient receptor potential vanilloid type 1 (TRPV1) receptor [85] (Fig. 3B). As a member of the thermosensitive TRP channel family, TRPV1 responds to thermal, mechanical, and chemical stimuli. Its activation leads to sensory neuron depolarization and the subsequent release of CGRP and substance P (SP), which



Fig. 3 Mechanisms by which tumor cells promote cancer pain: A. Tumor cells release NGF, which acts on TrkA receptors on sensory neurons, activating them through the RAS/MAPK/ERK pathway and transmitting pain signals. B. Tumor cells release inflammatory cytokines (e.g., TNF-a, IL-1, IL-6) and neurotrophic factors (e.g., NGF), targeting TRPV1 receptors on sensory neurons and promoting the secretion of pain-inducing substances such as SP and CGRP by these neurons. C-E. Tumor cells also influence sensory neurons through the secretion of miRNAs. C. Tumor cells release less miR-124, which allows for increased expression of the *Synpo* gene, promoting synaptic signal transmission and enhancing pain sensitivity. D. Tumor cells release miR-1a-3p, which acts on neurons to silence the *Clcn3* gene, increasing the sensitivity of dorsal root ganglia. E. Tumor cells secrete miR-21, miR-34a, and miR-324, leading to the reprogramming of sensory neurons into adrenergic neurons. The activation of endogenous pain-related genes in these neurons can intensify the pain experienced by cancer patients

facilitate pain transmission [86, 87]. For instance, models of bone cancer pain, TRPV1 is activated not only by metabolic byproducts such as formaldehyde but also by inflammatory mediators including tumor necrosis factor α (TNF- α), Interleukin-1 (IL-1), and IL-6, all of which are secreted by cancer cells [88, 89] (Fig. 3B). This activation enhances the sensitivity of dorsal root ganglia (DRG) neurons, contributing to the perception of pain [90]. Notably, in a melanoma model, either TRPV1 knockdown or CGRP receptor

inhibition significantly impaired tumor growth and increased survival in mice bearing B16F10 melanoma cells. Furthermore, TRPV1 expression across various immune cell types suggests a potential role in antitumor immune responses, a topic warranting further investigation [91, 92].

MicroRNAs (miRNAs) have emerged as critical regulators of neuropathic pain, influencing neuronal development, plasticity, and function [93-96]. In cancer, miRNAs packaged into tumor-derived extracellular vehicles (EVs) are differentially expressed and can modulate both the tumor microenvironment and neural signaling pathways (Fig. 3C-E). In a bone cancer pain model, a marked downregulation of miR-124 was detected in the spinal cord, correlating with enhanced pain signal transmission from the spinal cord to the brain [97] (Fig. 3C). This effect is partly mediated by miR-124's suppression of Synaptopodin (Synpo), a key protein in synaptic transmission. Meanwhile, miR-1a-3p was shown to exacerbate pain sensitivity in sensory neurons of the DRG by targeting the Clcn3 chloride channel gene [98] (Fig. 3D), highlighting the functional impact of specific miRNAs in pain regulation. This underscores the potential of miRNAs as therapeutic targets and biomarkers in cancer pain management. The aforementioned experimental findings indicate that miRNAs may serve as intermediaries in tumor-nerve interactions, modulating the transmission of cancer pain signals. Amit et al. reported significant findings in a p53-deficient oral squamous cell carcinoma (OCSCC) mouse model, where the expression of extracellular vesicles harboring specific miRNAs, namely miR-21, miR-34a, and miR-324, was diminished [99] (Fig. 3E). This reduction in miRNA-enriched extracellular vesicles from cancer cells was observed to trigger a restructuring of the existing sensory neurons, facilitating their transformation into adrenergic nerves. The activation of endogenous pain-related genes within these sensory neurons, such as *Ntrk2*, *Tac1*, and *Plcg1*, was identified as a mechanism that intensifies the experience of cancer pain [99].

In summary, cancer-associated pain results from a complex interplay of tumor-derived signals, immune responses, and neuronal adaptations. While inflammation plays a contributory role, a significant portion of cancer pain is neuropathic in origin, driven directly by cancer cells through the release of neuropeptides, the activation of cation channels such as TRPV1, and the modulation of neural gene expression by miRNAs. Together, these molecular and cellular events constitute a multifaceted mechanism through which tumors induce and sustain pain. Understanding these pathways not only offers insights into the biology of cancer pain but also provides promising targets for its effective management.

Cancer-nervous system related pathways

The interaction between the nervous system and tumors is multifaceted and dynamic, contributing significantly to cancer progression. With growing insight into the cancer-promoting effects of neural activity, an increasing number of signaling pathways linking the nervous system to tumor development have been identified and experimentally validated. These pathways play critical roles in modulating tumor cell behavior, including proliferation, invasion, and metastasis. In this section, we summarize key nervous system-related pathways that have been elucidated in recent years (Table 2).

Cholinergic receptor-related pathways

Cholinergic receptors, membrane-bound proteins that bind acetylcholine, are widely expressed throughout the central and peripheral nervous systems. Based on their pharmacological and molecular properties, they are broadly categorized into two major groups: nicotinic cholinergic receptors (nAChRs) and muscarinic cholinergic receptors (mAChRs). A growing body of evidence suggests that these receptors play critical roles in mediating nervous system-driven tumorigenesis and metastasis (Fig. 4A).

mAChRs are classified into five subtypes, M1 to M5, encoded by the genes cholinergic receptor muscarinic 1 (CHRM1) to cholinergic receptor muscarinic 5 (CHRM5). Each subtype contributes uniquely to neuraltumor signaling loop. Recent studies have confirmed that in PDAC mice, muscarinic agonists can rescue cancer progression in vagotomized mice. Specifically, activation of CHRM1 inhibits the epidermal growth factor receptor (EGFR)/MAPK/PI3K/AKT signaling cascade. Enhanced cholinergic signaling also suppresses cancer stem cell (CSC) populations, CD11b⁺ myeloid infiltration, TNF- α production, and hepatic metastatic growth, ultimately prolonging survival in PDAC-bearing mice [100]. In gastric cancer, Yu et al. demonstrated that tumor cells are capable of synthesizing and secreting acetylcholine, leading to autocrine and paracrine activation of muscarinic acetylcholine receptor 3 (M3R), which induces gastric cancer cell proliferation through trans-activation of EGFR signaling [101]. Furthermore, cholinergic signaling in Dclk1⁺ cell clusters within the gastric epithelium have been shown to induce NGF expression, which in turn stimulates cholinergic neuron growth, forming a positive feedback loop that sustains tumor-nerve interactions [102]. Similar tumor-promoting effects of M3R have been observed in pancreatic and colorectal cancers

Table 2 Cancer-nervous system related pathways

Neurosecretory factors	Receptors on tumor cells	Related pathways	Tumor changes
Acetylcholine	CHRM1	EGFR/MAPK/PI3K/AKT [100]	Inhibits cancer progression, reduces inflam- mation
	CHRM1 (in stroma) [54]	Unspecified	Promotes prostate tumor growth and metas- tasis
	CHRM3	EGFR transactivation [101]	Promotes gastric cancer proliferation
		NGF induction, cholinergic neuron growth [102]	Establishes tumor-nerve feedback loop
	CHRM4	AKT [54]	Induces neuroendocrine differentiation and invasion
	CHRNA5	$Ca^{2+}/CaMKII/GSK-3\beta/\beta$ -catenin [43]	Enhances β -catenin, BDNF expression, metastasis, drug resistance
Catecholamines	ADRB2	ETV1/c-KIT/MAPK [106–108]	Promotes tumorigenesis in GIST
		AKT/p53 [110]	Inhibits apoptosis, supports survival
		AKT/Beclin1/VPS34/HIF-1a [112]	Impairs autophagy, enhances resistance to stress
	ADRB3	Epac/JNK/BDNF [26, 121]	Promotes neuronal recruitment and tumor progression
		Epac/JNK/mTOR [113]	Stimulates ribosome biogenesis, cell cycle entry
NGF	TrkA	MAPK/ERK [122]	Promotes tumor progression
		EphA2/SRC [57]	Promotes tumor progression
		HIF-1a [123]	Promote tumor new angiogenesis, promote tumor growth
BDNF	TrkB	ERK/MAPK [124, 125]	Promote cell proliferation, inhibit cell apoptosis and stimulate angiogenesis
		PI3K/AKT/mTOR [126, 127]	Regulates cell proliferation, growth, size, metabolism, and movement
		PLC-γ/DAG [128]	Increase the survival rate of tumor cells and promote invasion
		Jak/STAT [129, 130]	Increase the survival rate of tumor cells and promote invasion
	p75 NTR	NF-κB/MMP9 [131]	Promote tumor cell survival and metastasis
		AKT/ERK [132]	Promote tumor cell survival and metastasis
Glutamate	NMDAR	FMRP/HSF1 [119, 120, 133]	Promotes invasion and metastasis via transla- tional regulation of stress response proteins
		CaMKII/MAPK/CREB, METTL3/m6A/HK2 [117, 118]	Enhances glycolysis, supports perineural inva- sion
	AMPAR	MAPK [134, 135]	Promotes invasion and migration in pancreatic cancer
		K-Ras/p38, K-Ras/p44/42[120]	Increases aggressiveness of precancerous lesions

[103]. In prostate cancer, acetylcholine released by parasympathetic nerve endings activates CHRM1 in mesenchymal stromal cells, promoting tumor proliferation and metastasis. Additionally, CHRM4 has been implicated in neuroendocrine differentiation, a critical step in prostate cancer progression. NGF-induced CHRM4 expression activates AKT signaling, which promotes cancer cell migration, invasion, and dissemination [54].

nAChRs are extensively involved in nerve-tumor interactions. Subunits such as α 3, α 4, α 5, and α 7 have been strongly associated with PDAC stem cell proliferation and resistance to inhibitory signals from neurotransmitters like GABA [104]. Selective knockdown of $\alpha 2/$ $\beta 4nAChR$ inhibits GABA production in PDAC cell lines and benign pancreatic epithelial cells [105]. Recent studies also implicate the acetylcholine/CHRNA5 ($\alpha 5$ -nAChR subunit) axis in promoting intrahepatic cholangiocarcinoma (ICC) metastasis. Acetylcholine binds CHRNA5, activating the Ca²⁺-dependent CaMKII/GSK-3\beta/βcatenin signaling pathway. This pathway upregulates β -catenin expression, enhances neural infiltration by



Fig. 4 Cancer-nervous system related pathway A. Cholinergic receptor-related pathways: CHRM1 activation suppresses EGFR/MAPK/PI3K/AKT signaling, inhibiting tumor progression. Conversely, tumor-derived acetylcholine activates M3R in an autocrine/paracrine manner, leading to EGFR transactivation and tumor proliferation. Cholinergic signaling also induces NGF expression, stimulating cholinergic nerve growth and forming a positive feedback loop. NGF upregulates CHRM4, activating AKT to promote invasion and metastasis. Acetylcholine binding to CHRNA5 triggers CaMKII/GSK-3B/B-catenin signaling, enhancing metastasis. B. Adrenergic receptor-related pathways: Neuronal signals activate ADRB receptors on tumor cells. ADRB2 promotes ETV1/c-KIT signaling via ERK and inhibits autophagy through AKT activation. ADRB3 promotes proliferation via Epac/JNK and mTOR pathways and stimulates BDNF release, enhancing neuronal support within the tumor microenvironment. C. Glutamate receptor-related pathways. Neuronal glutamate activates NMDAR on tumor cells, triggering calcium influx and downstream activation of FMRP/ HSF1, CaMKII/MAPK pathways, promoting tumor growth and progression. Glutamate also activates AMPAR, inducing MAPK, K-ras signalings, further enhancing tumor proliferation. D. Netrin-1/DCC-related pathway: Tumor-secreted netrin-1 binds neuronal DCC receptors, inducing FAK, Rac1, and Cdc42 phosphorylation, promoting nerve fiber growth. It also recruits PITPa to DCC, enhancing PI(5)P hydrolysis and neurite extension. E-H. Neuro-immune-cancer pathways. E. Norepinephrine activates ADRB2 on TAMs, inducing IL-33 and M2 polarization. TAMs secrete bFGF, stimulating Schwann cells via PI3K/AKT and promoting perineural invasion. F. ADRB2 on Tregs induces cAMP/PKA signaling, enhancing Treg migration and suppressive function; TGF-β/Smad2/3 promotes Treg differentiation, inhibiting anti-tumor immunity. G. Adrenaline enhances tumor exosome release and SP1 expression, activating neutrophils to secrete IL-1β, which triggers EGFR/PI3K/AKT and MEK/ERK pathways, facilitating EMT and metastasis. H. CGRP from nociceptors activates CAFs via cAMP/PKA, reducing IL-15 and impairing NK cells. CGRP also increases NGF, reinforcing immunosuppression and neuroinvasion

increasing BDNF, and promotes both metastasis and drug resistance in ICC [43].

In conclusion, cholinergic receptors, particularly the M3R and α 7-nAChRs, play integral roles in tumor progression by promoting proliferation, regulating inflammation, modulating the tumor microenvironment, and facilitating neuroinvasion. Targeting cholinergic signaling may offer promising therapeutic avenues in oncology.

Adrenergic receptors -related pathways

Beta-adrenergic receptors (ADRB), especially ADRB2 and ADRB3, are commonly found on the surface of various tumor cells and serve as critical mediators of neural influence on tumor biology. Under conditions of chronic stress, catecholamines such as norepinephrine are secreted by the sympathetic nervous system, activating ADRBs and modulating downstream signaling pathways involved in tumor progression. These downstream pathways serve as intermediaries through which the nervous system modulates tumors, bridging the influence between neurons and tumors (Fig. 4B).

ADRB2 is a principal mediator of catecholaminergic effects in tumors and plays a crucial role in the neural regulation of gastrointestinal stromal tumors (GIST). It promotes the ETS translocation variant 1 (ETV1)/c-KIT axis, a pathway essential for GIST development. ETV1, a lineage-specific transcription factor in GIST, facilitates tumorigenesis via MAPK-ERK signaling activated by ADRB2 stimulation [106–108]. Chronic restraint stress has been shown to trigger sustained catecholamine release, which promotes gastric epithelial transformation and carcinogenesis via ADRB2 signaling [109]. Local neuronal release of catecholamines also enhance AKT activation, promoting ubiquitination and degradation of p53, thereby inhibiting epithelial apoptosis and supporting tumor cell survival [110]. In addition, ADRB2-mediated upregulation of AKT disrupts the Beclin1/VPS34/ Atg14 complex, impairing autophagy and stabilizing HIF-1 α . This metabolic reprogramming enhances tumor cell resistance to hypoxia and other stressors, favoring survival in adverse microenvironments [111, 112]. Beyond ADRB2, ADRB3 contributes to neural-tumor communication under prolonged adrenergic stimulation. ADRB3 activation induces BDNF expression in tumor cells via an Epac (exchange protein activated by cAMP)/ JNK (c-Jun N-terminal kinase)-dependent pathway. BDNF is subsequently secreted into the tumor microenvironment, where it supports further neuronal recruitment and plasticity. While the relationship between Epac and JNK in this context, whether cooperative or parallel, remains to be fully clarified, both are implicated in regulating BDNF-mediated tumor progression. Moreover, ADRB3 signaling engages the mammalian target of rapamycin (mTOR) pathway to stimulate ribosome biogenesis, thereby facilitating G0-G1 cell cycle progression and impeding differentiation into adipocyte-like phenotypes [113]. In summary, ADRB2 and ADRB3 are central mediators of adrenergic signaling in tumors. By regulating cell proliferation, metabolic adaptation, immune evasion, and neural remodeling, these receptors constitute promising targets for therapies aimed at disrupting the neural regulation of tumor progression.

Glutamate receptor-related pathways

Glutamate, a key excitatory neurotransmitter, is predominantly active within the central nervous system (CNS), where it plays a crucial role in synaptic transmission and neuronal plasticity. Its receptors, including NMDAR and α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors (AMPAR), are essential mediators of CNS functions such as learning, memory, and neural communication [114–116]. Interestingly, emerging evidence suggests that glutamate also exerts regulatory effects beyond the CNS, particularly on peripheral tumors such as PDAC. These effects are mediated through the glutamate receptors expressed on tumor cells, including NMDAR and AMPAR, suggesting a potential role for glutamate signaling in cancer biology [117–120] (Fig. 4C).

One mechanistic link involves guanylate kinase-associated protein (GKAP), a scaffold protein of NMDARs, promotes fragile X mental retardation protein (FMRP)/ heat shock factor 1 (HSF1) signaling, which collectively drives tumor invasion and malignancy. Calcium influx through NMDARs enhances FMRP translation, and although the exact mechanism remains unclear, strong associations have been identified in candidate gene studies. FMRP binds HSF1 mRNA, promoting its translation and phosphorylation at serine 326, thereby activating HSF1. Intriguingly, FMRP also binds mRNAs encoding NMDAR subunits (GluN1, GluN2 A, GluN2B), suggesting a positive feedback loop that amplifies NMDAR signaling. Their reciprocal regulation appears complex and bidirectional, requiring further investigation [119]. In parallel, NMDAR-mediated calcium influx activates the CaMKII/MAPK pathways. This activation results in phosphorylation of CREB at Ser133, enhancing its transcriptional activity [118]. Notably, CREB activation induces Methyltransferase Like 3 (METTL3) expression, which upregulates hexokinase 2 (HK2) via N6-methyladenosine (m6A) mRNA modification, enhancing glycolysis and promoting PNI in tumor cells [117]. These pathways have been validated in mouse models of PDAC. Glutamate-mediated AMPAR activation in PDAC promotes tumor progression via the MAPK/K-ras pathway, supported by in vitro studies, while in vivo evidence remains limited [120] (Fig. 4C). These insights underscore the intricate role of glutamate receptor-related pathways in mediating neural modulation of tumor behavior, highlighting its potential as a therapeutic target in cancer treatment.

Netrin-1/DCC-related pathways

Netrin-1, a well-characterized neurotrophic factor, plays essential roles in the development of the central nervous system, spinal cord, and peripheral nerves. Beyond its physiological functions, it is increasingly recognized as a key modulator in tumor biology, particularly in relation to tumor-associated pain. Elevated expression of Netrin-1 within the tumor microenvironment activates specific neuronal receptors and initiates downstream signaling cascades that promote neural innervation of tumors, thereby contributing to the onset and persistence of cancer-related pain [136]. Emerging evidence highlights a strong association between Netrin-1 expression and tumor aggressiveness. Its upregulation has been documented across a broad range of malignancies, where it facilitates tumor-nerve interactions. Tumors may exploit Netrin-1 signaling to modulate neuronal activity, which in turn regulates both tumor behavior and immune responses [136, 137]. This bidirectional communication between the nervous system and tumors creates a mutually supportive environment that enhances tumor survival, progression, and pain generation (Fig. 4D).

DCC, a transmembrane receptor primarily expressed on neurons, serves as a principal receptor for Netrin-1. Activation of DCC has been shown to direct axonal growth through specific intracellular pathways. In models of bone cancer, DCC-mediated signaling contributes to increased innervation by CGRP⁺ sensory nerve fibers, exacerbating nociceptive signaling and cancer-induced bone pain [138]. When DCC on neurons is attracted and activated by Netrin-1 from cancer cells, it promotes phosphorylation of focal adhesion kinase (FAK) and subsequently Rac family small GTPase 1/cell division cycle 42 (Rac1/Cdc42) signaling pathway. This Netrin-1/ DCC/FAK/Rac1/Cdc42 axis drives the sprouting and elongation of peripheral CGRP⁺ nerve fibers, thereby enhancing nociceptive innervation during cancer pain progression [139]. These findings suggest that activation of the Netrin-1/DCC pathway in the tumor milieu facilitates neuroplastic changes that amplify pain perception. In addition to its role in nociceptive innervation, Netrin-1 also promotes axonal outgrowth through another mechanism involving phosphatidylinositol transfer protein- α (PITP α). Upon stimulation by Netrin-1, PITP α is recruited to the plasma membrane via interaction with DCC, which may induce conformational changes that enhance hydrolysis of phosphatidylinositol 5-phosphate (PI(5)P). This process likely contributes to axon elongation and may further promote local PI(5) P synthesis, reinforcing the signaling cascade. Although the full role of the Netrin-1/DCC/PITP α /PI(5)P pathway in the context of tumor-nerve interactions remain to be fully elucidated, it presents a promising mechanism by which tumors may hijack neuronal growth processes.

Importantly, Netrin-1 not only functions as a common neurotrophic factor guiding neuronal growth but also critically determines neuronal survival. Studies have shown in regions of the nervous system where local Netrin-1 is absent, axonal terminals and growth cones undergo morphological collapse reminiscent of apoptosis. This effect may not result solely from lack of downstream pathway activation but could also involve the unliganded DCC receptor, which has been suggested to exert pro-apoptotic effects in the absence of Netrin-1 binding. This mechanism, however, remains hypothetical and warrants further investigation [140].

In summary, Netrin-1/DCC signaling pathways are central to both normal neuronal development and pathological processes associated with cancer, particularly tumor-induced neuroplasticity and pain. Elucidating these mechanisms enhances our understanding of the tumor-nervous system interface and may inform novel therapeutic strategies aimed at alleviating cancer-related neurological complications.

Neuro-immune-cancer pathways

Neuronal activity has been increasingly recognized as a modulator of immune system function, which in turn plays a pivotal role in tumor development and progression. The immune system serves as a critical intermediary in the bidirectional communication between the nervous system and tumors, facilitating indirect neurotumoral interactions. Neural influences on immune cells may impair their ability to detect and eliminate tumor cells, thereby fostering tumor immune evasion. Consequently, the nervous system, via its regulation of immune components, can enhance tumor growth, progression, and metastasis. The following sections categorize these regulatory interactions according to immune cell types involved (Table 3).

Tumor-Associated Macrophages (TAMs)

TAMs are integral components of the tumor microenvironment (TME), and their phenotypes are dynamically regulated by neural signals [141, 142] (Fig. 4E). In particular, sympathetic nervous system activity modulates TAM behavior through the release of norepinephrine, which binds to β 2-adrenergic receptors (ADRB2) on TAMs. This interaction promotes the expression of immunosuppressive and pro-tumorigenic factors such as

Table 3 The changes	n immune cells within the tumor imm	une microenvironment influenced by th	e nervous system	
Immune cell types	Mechanism of action of the nervous system	Changes in immune cells	Effects on tumors	Key molecule
Tumor-Associated Mac- rophages (TAMs) [141]	Activate ADRB2 on tams [143]	Upregulate immunosuppressive molecules, pro-angiogenic VEGF [144], suppress pro-inflammatory cytokines	Impaire antitumor immunity, drive tumor progression [145]	IL-10↑, ARG1↑, VEGF↑, TNFa↓, IL-1β↓, IL-12↓
	Activate Schwann cells, upregulate GFAP	Activate Schwann cells, polarize into M2 turmor-promoting phenotypes, form a TAM- Schwann cell positive feedback loop	Accelerate tumor metastasis [146]	GFAP↑
	Secrete NGF, activate the TrkA receptor	Trigger PI3K/AKT pathway, promote mTOR activation, increase VEGF and IL-6	Enhance angiogenesis and tumor cell survival [161]	VEGF1, IL-61
	Secrete GDNF, activate the RET tyrosine kinase receptor	Regulate tumor cell metabolic repro- gramming, cytoskeletal dynamics, and microenvironmental adaptation	Promote tumor cells' migration enhances perineural invasion capabilities [162]	GDNF1
Regulatory T Cells (Tregs)	Activate the camp/PKA pathway in Tregs, promote their migration into the tumor microenvironment	Suppress T-cell proliferation, downregu- late cyclin D/E, upregulate p21/p27	Suppress the antitumor immune response, enhance their own prolifera- tion, reinforce immunosuppression, facili- tate tumor immune evasion [151]	TGF-β↑, cell cycle proteins↓, cell cycle inhibitors↑
	Activate the cAMP/PKA pathway in Tregs, promote their migration into the tumor microenvironment	Activating Foxp3, drive naïve CD4 ⁺ T-cell differentiation into immunosuppressive iTregs	Suppress the antitumor immune response, enhance their own prolifera- tion, reinforce immunosuppression, facili- tate tumor immune evasion [151]	TGF-B1, Foxp31
	Activate the cAMP/PKA pathway in Tregs, promote their migration into the turnor microenvironment	Amplify by inducing IL-10, IL-35, and TGF-β secretion, form a self-reinforc- ing immunosuppressive loop	Suppress the antitumor immune response, enhance their own prolifera- tion, reinforce immunosuppression, facili- tate tumor immune evasion [151]	IL-10f, IL-35 f, TGF-β†
	Activate the cAMP/PKA pathway in Tregs, promote their migration into the turnor microenvironment	Bind CD80/CD86 on APCs, recruite SHP2 phosphatase, inhibit TCR signaling and block T-cell activation, reinforce immune suppression	Suppress the antitumor immune response, enhance proliferation, reinforce immunosuppression, facilitate tumor immune evasion [151]	SHP2 phosphatase1
	Release NE, activate the (32-adrenergic receptor ((32-AR)	Trigger the camp-PKA signaling pathway, promote the secretion of IL-10 and TGF- β	Inhibit effector T cell activity, suppress anti-tumor immune responses [163]	lL-101, TGF-β secretion1
Neutrophils	Undergo enhanced production and modification, increase SP1 levels, facilitate neutrophil activation, secrete IL-1β	Activate MMP/ADAM proteases, release EGFR ligands, trigger EGFR-PI3K/AKT signaling [155], inhibit GSK-3ß, stabilize EMT transcription factor, downregu- late E-cadherin, upregulate vimentin, enhance cell motility	Enable superior adaptation to metastatic microenvironments, potentiate the met- astatic capacity of cancer cells [157]	lL−1β1, EGFR↑
	Undergo enhanced production and modification, increase SP1 levels, facilitate neutrophil activation, secrete IL-1ß	Bind to IL-1R, activate the MEK/ERK pathway [156], induce the expression of mesenchymal genes, enhance cell migration and invasion capabilities	Enable superior adaptation to metastatic microenvironments, potentiate the met-astatic capacity of cancer cells [157]	lL-1βî
	Secrete SPP1, release catecholamines by sympathetic nerves, enhance CXCL1 secretion, CXCL1 binds to the CXCR2 receptor	SPP1 binds to neutrophil surface receptor, activate the ERK pathway, promote NET formation; nets capture circulating tumor cells, release inflammatory mediators	Promote tumor cell survival and invasion [164]	SPP11, inflammatory mediators1

Table 3 (continued)				
Immune cell types	Mechanism of action of the nervous system	Changes in immune cells	Effects on tumors	Key molecule
Natural Killer (NK) Cells	Release CGRP, bind to the CGRP receptors RAMP1 and CALCRL, activate AC, elevate intracellular camp levels, stimulate PKA [158]	Phosphorylate the transcription factor CREB, suppress IL-15 [159]	Drive tumor progression, exacerbate cancer-related pain, promote PDAC aggressiveness and pain hypersensitiv- ity [160]	CGRP↑, IL-15↓
	Release CGRP, bind to the CGRP receptors RAMP1 and CALCRL, activate AC, elevate intracellular camp levels, stimulate PKA [158]	Secret NGF, bind to the TrkA receptor, enhance CGRP release, establish a self- reinforcing feedback loop	Drive tumor progression, exacerbate cancer-related pain, promote PDAC aggressiveness and pain hypersensitiv- ity [160]	CGRP↑
	Induce tumor cells or antigen-presenting cells to express PD-L1	Bind to the PD-1 receptor, activate down- stream inhibitory signaling pathways, suppress the PI3K/AKT/mTOR pathway, reduce the synthesis and release of cyto- toxic granules	Suppres anti-tumor immunity [165]	PD-L11, cytotoxic granules

interleukin-10 (IL-10), arginase 1 (ARG1), which inhibits T cell metabolism [143], and vascular endothelial growth factor (VEGF, which promotes angiogenesis) [144]. Simultaneously, norepinephrine suppresses TAM production of pro-inflammatory cytokines such as TNF- α , IL-1 β , and IL-12, thereby weakening anti-tumor immune responses and facilitating tumor progression [145]. Moreover, TAMs secrete basic fibroblast growth factor (bFGF), which binds to bFGF receptors on Schwann cells, activating the PI3K-Akt signaling pathway. Akt then phosphorylates and activates c-Myc, which translocates to the nucleus to upregulate glial fibrillary acidic protein (GFAP), a marker of Schwann cell activation. These activated Schwann cells promote PNI by increasing migration and secreting interleukin-33 (IL-33), which recruits additional macrophages and further induces M2 polarization. This positive feedback loop between TAMs and Schwann cells plays a central role in the neural invasion and metastatic behavior of PDAC [146]. Overall, interactions between neurons, glial cells, and TAMs, whether through neural modulation of cytokine profiles or reciprocal activation loops, serve to suppress antitumor immunity while enhancing tumor progression and dissemination.

Regulatory T Cells (Tregs)

Tregs in the tumor microenvironment predominantly exhibit immunosuppressive effects against anti-tumor immunity [147]. With advancing research, increasing evidence suggests that this suppression is intricately linked to neural mechanisms. Specifically, the nervous system regulates the proliferation, metabolism, and immunosuppressive functions of Tregs through neurotransmitters, nerve fiber infiltration, and neuro-immune signaling pathways, thereby promoting tumor immune evasion [148]. Sympathetic nerve-released norepinephrine also influences the behavior of regulatory T cells (Tregs) by activating the cAMP/PKA signaling pathway through β 2-adrenergic receptors. This enhances Treg migration into the TME, where they execute potent immunosuppressive functions (Fig. 4F). Tregs inhibit anti-tumor immunity through multiple mechanisms: On the one hand, Tregs secrete TGF-B, which downregulates cell cycle proteins (e.g., cyclin D/E) and upregulates cell cycle inhibitors (e.g., p21, p27) via the Smad2/3 signaling pathway, inducing G1 phase arrest and inhibiting T cell proliferation [149]. Besides, TGF-β activates Foxp3, a key transcription factor in Tregs, driving the differentiation of naive CD4⁺ T cells into induced Tregs (iTregs). TGF- β also further amplifies immunosuppression through Smad3 signaling, promoting Treg secretion of IL-10, IL-35, and additional TGF- β [150], which forms a positive feedback loop. On the other hand, Tregs express high levels of CTLA-4, which binds to CD80/CD86 on antigen-presenting cells (APCs), leading to recruitment of the SHP2 phosphatase, suppression of TCR signaling, and blockade of T cell activation [151]. Through these pathways, Tregs not only dampen anti-tumor immune responses but also promote their own expansion, thereby reinforcing immune suppression and facilitating tumor immune evasion [151].

Neutrophils

Neutrophils are often regarded as a "double-edged sword" in anti-tumor immunity, exhibiting distinct roles across different tumor microenvironments [152]. The nervous system significantly modulates the dual roles of neutrophils in tumor progression, both pro-tumorigenic and anti-tumorigenic, through neurotransmitter release, microenvironment remodeling, and direct cellular interactions [153, 154] (Fig. 4G). Under chronic stress conditions, chronic stress affects the activity of the sympathetic nervous system and activates β -adrenergic receptors on tumor cells, which induces the increased SP1 levels in tumor-derived exosomes (TDES) facilitate neutrophils activation by binding to TLR4 on neutrophils and activates the TLR4/NF-KB signaling pathway, leading to IL-1 β secretion of neutrophils. Subsequently, IL-1 β promotes cancer metastasis via coordinated activation of both EGFR-dependent PI3K/AKT and IL-1R-dependent MEK/ERK signaling pathways in cancer cells [155, 156].

As for the EGFR-dependent PI3K/AKT pathways, IL-1 β activates Matrix metalloproteinase (MMP)/ ADAM proteases to release EGFR ligands, resulting in EGFR phosphorylation and subsequent PI3K/AKT pathway activation. This pathway inhibits GSK-3 β activity to stabilize EMT transcription factors, which subsequently downregulates E-cadherin expression, promotes cytoskeletal reorganization, and upregulates vimentin expression, finally leads to the enhancement of cell motility and result in promoting cancer metastasis. Meanwhile, IL-1 β can also directly binds to IL-1R on tumor cells and activates the MEK/ERK pathway through the IKK β /Tpl2 cascade, inducing the expression of mesenchymal genes and enhancing cell migration and invasion capabilities.

This dual-pathway synergistic mechanism originated from IL-1 β secreted by neutrophils under the influence of neuroendocrine exosomes drives epithelial cells to retain partial epithelial markers through incomplete E-cadherin loss while simultaneously gaining mesenchymal characteristics via vimentin upregulation, thereby establishing a hybrid epithelial-mesenchymal phenotype. Such phenotypic plasticity enables superior adaptation to metastatic microenvironments and ultimately potentiates the metastatic capacity of cancer cells [157].

Natural Killer (NK) cells

NK cells are key effectors in anti-tumor immunity. However, their function can be suppressed through neuroimmune interactions within the TME (Fig. 4H). In the TME of PDAC, nociceptor neurons release CGRP, which binds to the CGRP receptors RAMP1 and CALCRL on the surface of cancer-associated fibroblasts (CAFs). Upon receptor binding, CGRP activates adenylate cyclase (AC) via the Gs protein, elevating intracellular cAMP levels and stimulating PKA [158]. The cAMP/PKA pathway phosphorylates the transcription factor CREB, leading to transcriptional suppression of IL-15, a critical cytokine for the survival, proliferation, and activation of NK cells [159]. As a result, reduced IL-15 levels impair NK cell recruitment and effector function, compromising tumor immunosurveillance. Meanwhile, CGRP-stimulated CAFs secrete NGF, which activates TrkA receptors on nociceptor neurons, further enhancing CGRP release. This positive feedback loop exacerbates both immune suppression and nociceptor sensitization. This neuroimmune interplay not only facilitates immune evasion by suppressing NK cell activity but also contributes to neural remodeling and cancer pain, thereby promoting PDAC aggressiveness and hypersensitivity [160].

Implication of targeting neural-cancer interactions

The increasing recognition of neuro-tumor interactions has highlighted the pivotal role of the nervous system in tumor initiation, progression, and metastasis. Advancements in this area, supported by accumulating preclinical and translational studies, have unveiled novel therapeutic targets and strategies. Both pharmacological and nonpharmacological interventions are now being explored to modulate these interactions. A growing body of evidence supports the development of therapies aimed at disrupting neural inputs that facilitate tumor development, thus offering potential avenues to modulate disease progression. Moreover, several non-invasive therapies have already been translated into clinical practice, underscoring their therapeutic promise and clinical relevance.

Drug therapies targeting the neurobiology of cancer Targeting nervous system to inhibit tumor growth and metastatic spread

From a microscopic perspective, neuronal secretion of cell adhesion molecules, neurotrophic factors, and neurotransmitters can regulate tumor growth and metastasis, making the modulation of neuronal secretion a potential anti-tumor strategy (Fig. 5A-C, Table 4). As previously discussed, cholinergic receptor signaling mediated by acetylcholine is essential in driving cancer proliferation, invasion, and metastasis. Both pharmacological inhibition and genetic knockout of the muscarinic

acetylcholine M3 receptor have shown inhibitory effects on gastric, small intestine, and breast tumors. Botulinum toxin (BoNT), which blocks acetylcholine release, has also been shown to suppress tumor growth, suggesting a functional link between acetylcholine signaling and cancer progression, although the precise mechanism remains to be fully elucidated [166] (Fig. 5A).

Glutamatergic signaling also plays a key role in neurotumor communication. mGluR1, a metabotropic glutamate receptor, is required to maintain a tumorigenic phenotype, as shown by inducible RNA silencing systems. For instance, treatment of Michigan Cancer Foundation-7 (MCF-7) xenografts with the glutamate release inhibitor riluzole resulted in significant suppression of tumor progression [167, 168] (Fig. 5B). Neurotrophic factors such as BDNF and NGF further drive tumor growth by activating Trk receptors on tumor cells. Trk inhibitors like larotrectinib and entrectinib have shown significant and sustained efficacy in clinical trials, although resistance remains a challenge; next-generation Trk inhibitors are currently under development to address this issue [169] (Fig. 5C).

Denervation studies provide additional evidence for the contribution of the nervous system to tumor progression. Zhao et al. demonstrated that surgical vagotomy or localized BoNT-A injection significantly reduced tumor development in three distinct gastric cancer mouse models [14]. Likewise, Magnon et al. reported that both chemical and genetic ablation of β 2- and β 3-adrenergic signaling suppressed prostate tumor growth and metastasis in murine models [12]. Notably, optogenetic stimulation of the optic nerve has been shown to initiate gliomagenesis, whereas sensory deprivation (e.g., dark rearing) inhibited tumor development in vivo, highlighting the importance of neural activity in tumor initiation [170].

Despite compelling evidence from animal models, translation to human cancer therapy requires further investigation. Many neural-targeting interventions exhibit only transient efficacy, with tumors eventually adapting and circumventing these pathways. Thus, strategies that target neural influences must be integrated into broader, multimodal treatment approaches to achieve durable clinical benefits.

Remodeling the immune system via neural modulation for cancer therapy

The immune system also serves as a vital mediator in the interaction between the nervous system and tumors (Fig. 5D-F, Table 4). Neural regulation within the tumor microenvironment can dampen immune surveillance and promote tumor immune evasion. Therefore, targeting neural-immune crosstalk represents a promising strategy



Fig. 5 Implication targeting neural-cancer interaction: A. Botulinum toxin (BoNT) may inhibit tumor progression by suppressing neuronal acetylcholine release. B. An induced Grm1 silencing RNA system can block mGluR1, disrupting glutamate-mediated neural influences in the tumor microenvironment and exerting antitumor effects. Riluzole, a glutamate release inhibitor, has been shown to suppress breast cancer progression.
C. Trk inhibitors can suppress neuron-secreted neurotrophic factors such as BDNF and GNF, which promote tumor development by binding to Trk receptors on tumor cells. D. MDK inhibitors can block neuron-mediated CD8⁺ T-cell activation via MDK secretion, thereby preventing tumor immune evasion. E. Inhibiting the TGF-β pathway may suppress neuronal secretion of TGF-β, which otherwise promotes CD4⁺ T-cell proliferation and contributes to cancer progression. F. Blocking FasL interactions with neutrophils can reverse FasL-induced immunosuppressive effects on tumor-specific CD8⁺ T cells, thereby inhibiting tumor growth and progression

not only for tumor control but also for addressing tumorinduced neurological complications.

Neurons can influence immune cells through several mechanisms. For example, secretion of MDK can activate CD8⁺ T cells in a manner that paradoxically promotes immune evasion. In vitro studies by Liu et al. showed that MDK inhibitors enhanced the efficacy of immune modulators in ovarian cancer cells [171] (Fig. 5D). Neurons also secrete TGF- β to stimulate CD4⁺ T cell proliferation and produce Fas ligand (FasL) to induce T cell apoptosis [172, 173]. In basal-like breast cancer (BLBC), low TGF- β activity correlates with heightened CD4⁺ T cell function and reduced recurrence, suggesting that inhibition of TGF-β signaling could enhance immunotherapeutic outcomes [174]. These data indicate that inhibiting the TGF β pathway may serve as a promising therapeutic strategy to boost the immune system's efficacy in treating BLBC (Fig. 5E). Fas is a crucial factor in tumor immunotherapy. Shan et al. found that overexpressing FasL enabled neutrophils to acquire immunosuppressive functions against tumor-specific CD8⁺ T cells, promoting the growth and progression of human gastric cancer cells both in vitro and in vivo. This effect could be reversed by blocking FasL on these neutrophils [175] (Fig. 5F).

The nervous system also exerts regulatory effects directly within tumor cells. For instance, Wang et al. demonstrated that blocking acetylcholine signaling with the receptor antagonist 4-DAMP reduced PD-L1 expression and self-renewal in CD133⁺ thyroid cancer cells [29]. In a related pathway, the neurotransmitter GABA, acting through GABA_B receptors on tumor cells, has been shown to impair T cell infiltration. Sun et al. found that combining the GABA_B receptor agonist baclofen with anti-PD-L1 therapy enhanced immune response and tumor suppression in a breast cancer mouse model [176]. Collectively, these findings highlight the therapeutic

Table 4 Drugs targeting the Cancer-nervous system

Drug Name	Mechanism of Action	Cancer Type	Drug Type	Research Stage
Propranolol [177]	Blocks β2 receptors, inhibits sympathetic signaling, reduces inflammatory cytokines and pro-metastatic molecules	Breast cancer	β2-adrenergic receptor antagonist	Preclinical research
Carvedilol [178]	Blocked the effects of sym- pathetic nervous system activation	Breast cancer	β-adrenergic receptor antagonist	Retrospective clinical studies
Atenolol [179]	Inhibits sympathetic nerve- driven metabolic repro- gramming to reduce tumor proliferation	Breast cancer	β 1-adrenergic receptor antagonist	Preclinical research
Tiotropium [180]	Binds to M3 receptor to form a complex	Lung cancer	M3 muscarinic receptor antagonist	Retrospective clinical studies
Atropine [181]	Attenuates immunosup- pressive markers and M3R via inhibition of EGFR/AKT/ ERK signaling pathways	Colorectal cancer	Non-selective M1-M5 antagonist	Preclinical research
Larotrectinib [182]	Targets NTRK fusion genes, blocks MAPK/ERK and PI3K/ AKT pathways	NTRK fusion-positive solid tumors (e.g., Sarcoma)	TrkA/B/C inhibitor	Preclinical research
Entrectinib [182]	Inhibits Trk signaling and per- ineural invasion	NTRK fusion-positive solid tumors (e.g., Lung cancer)	Trk/ROS1/ALK inhibitor	Preclinical research
Selitrectinib [169]	Overcomes Trk kinase domain mutations	Trk kinase domain mutation- resistant tumors	Second-gen Trk inhibitor	Preclinical research
Repotrectinib [183]	Penetrates bloodbrain bar- rier to suppress intracranial metastasis	Breast cancer brain metas- tasis	Trk/ROS1/ALK inhibitor	Preclinical research
Cabozantinib [184, 185]	Decreases proliferation and migration of osteosar- coma cells, inhibiting ERK and AKT signaling pathways	Osteosarcoma	TrkB/MET/VEGFR2 inhibitor	Phase II clinical trial
SR48692 [186]	Inhibits MAPK/ERK, PI3K/AKT, and PKC pathways to sup- press tumor proliferation	Pancreatic cancer	Neurotensin receptor (NTSR1) antagonist	Preclinical research
Naltrexone [177, 187]	Activates TLR4 signaling to enhance immunity and induce apoptosis	Pancreatic cancer, breast cancer	Non-selective opioid antago- nist (low-dose)	Preclinical research
MDK inhibitor (iMDK) [171]	Inhibits MDK-mediated CD8 ⁺ T cell suppression	Ovarian cancer	Small molecule inhibitor	Preclinical research
TGFβ inhibitor [174]	Blocks TGFβ signaling to enhance CD4 ⁺ T cell activity	Basal-like breast cancer	Small molecule/antibody	Preclinical research
FasL blocker [175]	Blocks FasL-mediated T cell apoptosis	Gastric cancer	Antibody	Preclinical research
Baclofen [176]	GABA _B receptor agonist; stabilizes PD-L1	Breast cancer	Small molecule agonist	Preclinical research

potential of targeting neuro-immune interactions to enhance anti-tumor immunity. However, while immunotherapy has revolutionized cancer treatment, its efficacy remains variable, necessitating further refinement and the development of new immunotherapies specifically designed to disrupt neural-tumor immune pathways.

Non-invasive, non-pharmacological approaches related to the neurobiology of cancer

While pharmacological approaches targeting neural-cancer interactions have shown considerable promise, their clinical application is often limited by serious side effects, such as chemotherapy-induced peripheral neuropathy (CIPN) and neurotoxicity associated with immune checkpoint inhibitors (ICIs) [188, 189]. In recent years, non-invasive, non- pharmacological interventions have gained attention for their ability to prevent cancer progression, enhance the efficacy of existing treatments, and improve patient quality of life without inducing significant adverse effects [190–192].

Accumulating evidence over the past decade has established a clear link between psychological stress and cancer progression. These effects are largely mediated by the sympathetic-adrenal-medullary (SAM) axis and its catecholaminergic outputs, such as norepinephrine and epinephrine. These stress mediators promote tumor progression by stimulating cancer cell proliferation, angiogenesis, and motility; inhibiting cytotoxic immune cell activity; and enhancing the tumor-promoting functions of immune-suppressive cells. Furthermore, stress-induced sympathetic signaling has been shown to diminish the effectiveness of conventional treatments, including surgery, chemotherapy, radiotherapy, and immunotherapy [193]. Consequently, stress management and psychological interventions are now considered critical components of comprehensive cancer care. Psychotherapeutic strategies, particularly those targeting neuroendocrine stress responses, have demonstrated the ability to modulate sympathetic output, reduce excessive adrenergic signaling, and enhance immunochemotherapeutic efficacy [193, 194]. Among these, Cognitive Behavioral Therapy (CBT) has shown significant benefit. A meta-analysis that included 13 controlled trials revealed that CBT substantially improved psychological resilience in cancer patients [195]. Similarly, mindfulness-based and positive psychology interventions, which encourage present-moment awareness and nonjudgmental acceptance, have demonstrated efficacy in reducing distress and improving emotional well-being. A meta-analysis of 21 clinical studies confirmed their effectiveness in alleviating psychological burden among cancer survivors [196, 197].

Behavioral interventions such as regular exercise, meditation, and yoga have emerged as adjunctive therapies with neurobiological benefits relevant to cancer care. These activities are thought to modulate autonomic nervous system balance, particularly by enhancing vagal tone. Increased vagal activity has been associated with elevated BDNF levels, TrkB pathway activation, improved neurogenesis, and synaptic plasticity, all of which contribute to neuroimmune regulation and possibly tumor inhibition [198]. Importantly, vagal activity can be non-invasively assessed through heart rate variability (HRV), a biomarker of autonomic function. Higher HRV is associated with improved cancer prognosis, including lower tumor marker levels and reduced mortality. HRV biofeedback (HRV-B), a method involving slow, rhythmic breathing combined with visual feedback, can enhance vagal tone and suppress sympathetic activity. This not only improves immune responses, such as increased natural killer (NK) and CD8⁺ T cell activity, but may also reduce oxidative stress and tumor-promoting signaling within the tumor microenvironment [199]. These findings suggest that autonomic regulation through behavioral strategies may offer a safe, effective, and low-cost avenue for improving outcomes in cancer patients. Further investigation into the neurobiological mechanisms underlying these effects may help integrate such interventions more fully into standard oncology care.

Acupuncture and electroacupuncture (EA), cornerstone therapies in traditional Chinese medicine, have demonstrated efficacy in alleviating cancer-related symptoms, mitigating treatment side effects, and reducing cancer-related pain [200, 201]. Mechanistically, these therapies may activate mechanoreceptors and initiate afferent signaling through the ventrolateral fasciculus to specific brain nuclei. This activates descending pain inhibitory pathways involving endogenous opioid peptides, serotonin (5-HT), and other neurotransmitters [200]. Reflecting growing clinical support, the American Society of Clinical Oncology and the Society for Integrative Oncology issued a 2022 guideline recommending acupuncture or acupressure for the management of general cancer pain [202]. Notably, the IMPACT randomized controlled trial by Epstein et al. found that acupuncture combined with massage significantly alleviated pain and co-occurring symptoms (fatigue, insomnia) in patients with advanced cancer [203]. Transcutaneous electrical acupoint stimulation (TEAS), a non-invasive alternative to traditional acupuncture that uses surface electrodes, has also demonstrated efficacy in clinical settings. TEAS has been effective in relieving both acute and chronic postoperative pain in breast cancer patients, improving serum protein levels, and accelerating postoperative recovery in colorectal cancer cases In a study by Tian et al., TEAS significantly reduced postoperative pain scores in pancreatic cancer patients from day 1 to 4 postsurgery, reinforcing the analgesic potential of acupuncture-based modalities [204].

Compared with traditional therapies, which often suffer from drug side effects and limited efficacy of surgical methods, non-invasive and non-pharmacological therapies such as psychological interventions, behavioral therapy, and acupuncture offer several advantages: safety, accessibility, cost-effectiveness, and the potential to improve both tumor control and quality of life [193]. Crucially, many of these therapies exert their effects by modulating nervous system activity, thereby offering unique opportunities for integration into the emerging field of cancer neuromodulation. Future interdisciplinary research combining oncology, neurology, immunology, and behavioral science is essential to further elucidate these mechanisms and optimize such interventions. This integrated approach holds significant promise for enhancing survival and long-term well-being in cancer patients.

Conclusions and perspectives

Historically, the role of the nervous system in tumorigenesis, progression, and metastasis has been largely overlooked. However, recent advancements in cancer neurobiology have highlighted the central involvement of neural mechanisms in oncogenesis. Over the past decade, this emerging field has gained substantial traction, demonstrating that neural regulation plays a pivotal role in at least five of the eight hallmarks and two enabling characteristics of cancer. Cancer development induces local nerve remodeling, and nerve growth with secreted factors introduces novel dynamics in cancer tissue proliferation and invasion, resulting in a self-perpetuating cycle that escalates the complexity and peril of tumor progression [205]. Neurotrophic factors and neurotransmitters serve as the intermediaries in the crosstalk between cancer and the nervous system [68]. Elevated expression levels of neurotrophic factors in tumor cells can stimulate the ingrowth of nerves within the tumor microenvironment and modulate neuronal excitability [71, 206]. Furthermore, the nervous system exerts its influence on tumors primarily through the regulation of the immune system, hormonal levels, and electrical signaling. Recent studies have evidenced that central nervous system stress can disrupt leukocyte trafficking and the secretion of inflammatory cytokines [207, 208]. Beyond the secretion of neurotrophic factors, the neuron-like alterations in cancer cells present a compelling feature of the tumor's regulation of the nervous system [209]. Tumor aggressiveness is significantly associated with the neuron-like synapses of cancer cells. Tumor cells are capable of forming tumor-neuron synapses with neurons. Upon metastasis to the brain, breast cancer cells can engage with neuronal synapses, creating pseudotriadic synapses that foster tumor proliferation [210].

In conclusion, the intricate crosstalk between the nervous system and cancer has opened new and promising avenues for therapeutic intervention. Nevertheless, several challenges must be addressed to translate these findings into clinical practice. First, the lack of a clear distinction between cancer-associated nerves and normal neural tissues presents a major obstacle in designing targeted therapies that avoid off-target effects. Second, although animal models suggest that surgical or genetic denervation can suppress tumor growth and metastasis, it remains uncertain whether pharmacological neural blockade can replicate these effects safely and effectively. Third, the potential for integrating neuromodulatory strategies with existing cancer therapies, such as immunotherapy or chemotherapy, requires systematic evaluation to determine synergistic benefits. Future research should aim to delineate the molecular and functional differences between normal and cancer-associated nerves, establish reliable biomarkers for neural involvement in cancer, and explore multimodal treatment paradigms targeting both the tumor and its neural environment. A deeper understanding of cancer neurobiology will not only advance our knowledge of tumor pathophysiology but also facilitate the development of innovative, mechanism-based therapies that harness the nervous system as a therapeutic ally.

Abbreviations

ADRB	Beta-adrenergic receptors
AKT	Ak strain transforming
AMPAR	a-Amino-3-Hydroxy-5-Methyl-4-Isoxazolepropionic Acid Receptor
BDNF	Brain-Derived Neurotrophic Factor
BoNT	Botulinum toxin
CCL2	C-C motif chemokine ligand 2
CCI 4	C-C motif chemokine ligand 4
CCL5	C-C motif chemokine ligand 5
CGRP	Calcitonin gene-related peptide
CHRM1	cholinergic receptor muscarinic 1
CHRM3	cholinergic receptor muscarinic 3
CHRM4	cholinergic receptor muscarinic 4
CHRNA5	a5 nAChR subunit
CREB	CAMP Response Element-binding Protein
CXCL12	C-X-C motif chemokine ligand 12
CX3CL1	C-X3-C motif chemokine ligand 1
DCC	Deleted in colorectal cancer
EMT	Enithelial-mesenchymal transition
Enac	Exchange protein activated by cAMP
FRK	Extracellular signal-regulated kinase
FAK	Focal adhesion kinase
Fasl	Fas ligand
EMRP	Fragile X mental retardation protein
GABA	v-Aminohutvric Acid
GAP-43	Growth-associated protein 43
GIST	Gastrointestinal stromal tumors
GKAP	Guanylate kinase associated protein
GSK-3B	Glycogen Synthase Kinase 38
HSE1	Heat shock factor 1
	intrahenatic cholangiocarcinoma
INK	C-lun N-terminal kinase
mAChRs	
MAPK	Mitogen-Activated Protein Kinase
MDK	Midkine
mGluR1	Metabotropic glutamate receptor 1
MMP	Matrix metalloproteinase
nAChRs	nicotinic cholinergic receptors
NGE	Nerve Growth Factor
NK	Natural Killer
NMDAR	N-methyl-D-aspartate receptor
NPY	Neuropeptide Y
n75 NTR	P75 neurotrophin recentor
PDAC	pancreatic ductal adenocarcinoma
PD-L1	Programmed cell death 1 ligand 1
PI(5)P	Phosphatidylinositol 5-phosphate
PI3K	Phosphatidylinositol 3-kinase
PITPa	Phosphatidylinositol transfer protein-g
PNI	Perineural invasion
S4F	Semaphorin 4E

STAT3	Signal Transducer and Activator of Transcription 3
Synpo	Synaptic foot protein
TAMs	Tumor-Associated Macrophages
TAST	Tumor-Activated Schwann Cell Trajectories
TGFβ	Transforming growth factor beta
TIMP1	tissue inhibitor of matrix metalloproteinases 1
TM	Tumor microtubes
TME	tumor microenvironment
TNT	Tunneling nanotubes
Tregs	Regulatory T Cells
TrkB	Tropomyosin receptor kinase B
TrkC	Tropomyosin receptor kinase C
TRPV1	Transient receptor potential vanilloid type 1
Ttyh1	Tweety-homolog 1

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Authors' contributions

Sirui Huang and Jing Zhu were responsible for writing and revising the manuscript, creating the figures, and constructing the tables. Linglu Yu contributed to the figure editing and revision process. Yue Hu and Yan Huang offered funding support and played a role in the conception, design, editing, and supervision of the manuscript sections. All authors reviewed and agreed on the final version of the manuscript.

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Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Author details

¹School of Integrative Medicine, Nanjing University of Chinese Medicine, Nanjing 210023, Jiangsu, China. ²Department of Ultrasound, Nanjing Hospital of Chinese Medicine Affiliated to Nanjing University of Chinese Medicine, Nanjing 210022, China. ³Department of Neurology, Nanjing Hospital of Chinese Medicine affiliated to Nanjing University of Chinese Medicine, Jiangsu, Nanjing 210001, China. ⁴Shen Chun-Ti Nation-Famous Experts Studio for Traditional Chinese Medicine Inheritance, Changzhou TCM Hospital Affiliated to Nanjing University of Chinese Medicine, Jiangsu 213003, Changzhou, China.

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References

- Monje M, Borniger JC, D'Silva NJ, Deneen B, Dirks PB, Fattahi F, Frenette PS, Garzia L, Gutmann DH, Hanahan D, et al. Roadmap for the Emerging Field of Cancer Neuroscience. Cell. 2020;181:219–22.
- 2. Moore CH: Observations on the Division of the Gustatory Nerve, and on Ligature of the Lingual Artery in the Treatment of Cancer of the Tongue. Med Chir Trans 1862, 45:47–62.41.
- Finlayson J, Coats J. On a Case of Cancer of the Œsophagus Involving the Recurrent Laryngeal Nerve and the Body of the Last Dorsal Vertebra. Glasgow Med J. 1890;34:161–5.
- Fankhauser R, Luginbühl H, McGrath JT. Tumours of the nervous system. Bull World Health Organ. 1974;50:53–69.
- Spaulding WL. Glioma of the optic nerve and its management. Am J Ophthalmol. 1958;46:654–8.
- 6. Wardle EN. Nerve-sheath tumours. Br J Surg. 1957;45:58-61.
- Dodge HW Jr, Love JG, Craig WM, Dockerty MB, Kearns TP, Holman CB, Hayles AB. Gliomas of the optic nerves. AMA Arch Neurol Psychiatry. 1958;79:607–21.
- Mark GJ. Basal cell carcinoma with intraneural invasion. Cancer. 1977;40:2181–7.
- 9. Liebig C, Ayala G, Wilks JA, Berger DH, Albo D. Perineural invasion in cancer: a review of the literature. Cancer. 2009;115:3379–91.
- Vincent D, Dubas F, Hauw JJ, Godeau P, Lhermitte F, Buge A, Castaigne P. Nerve and muscle microvasculitis in peripheral neuropathy: a remote effect of cancer? J Neurol Neurosurg Psychiatry. 1986;49:1007–10.
- Adriaenssens E, Vanhecke E, Saule P, Mougel A, Page A, Romon R, Nurcombe V, Le Bourhis X, Hondermarck H. Nerve growth factor is a potential therapeutic target in breast cancer. Cancer Res. 2008;68:346–51.
- Magnon C, Hall SJ, Lin J, Xue X, Gerber L, Freedland SJ, Frenette PS. Autonomic nerve development contributes to prostate cancer progression. Science. 2013;341:1236361.
- Pundavela J, Roselli S, Faulkner S, Attia J, Scott RJ, Thorne RF, Forbes JF, Bradshaw RA, Walker MM, Jobling P, Hondermarck H. Nerve fibers infiltrate the tumor microenvironment and are associated with nerve growth factor production and lymph node invasion in breast cancer. Mol Oncol. 2015;9:1626–35.
- Zhao CM, Hayakawa Y, Kodama Y, Muthupalani S, Westphalen CB, Andersen GT, Flatberg A, Johannessen H, Friedman RA, Renz BW, et al: Denervation suppresses gastric tumorigenesis. Sci Transl Med 2014, 6:250ra115.
- Zhou Y, Shurin GV, Zhong H, Bunimovich YL, Han B, Shurin MR. Schwann Cells Augment Cell Spreading and Metastasis of Lung Cancer. Cancer Res. 2018;78:5927–39.
- 16. Gysler SM, Drapkin R: Tumor innervation: peripheral nerves take control of the tumor microenvironment. J Clin Invest 2021, 131.
- Silverman DA, Martinez VK, Dougherty PM, Myers JN, Calin GA, Amit M. Cancer-Associated Neurogenesis and Nerve-Cancer Cross-talk. Cancer Res. 2021;81:1431–40.
- Hanahan D, Monje M. Cancer hallmarks intersect with neuroscience in the tumor microenvironment. Cancer Cell. 2023;41:573–80.
- Balood M, Ahmadi M, Eichwald T, Ahmadi A, Majdoubi A, Roversi K, Roversi K, Lucido CT, Restaino AC, Huang S, et al. Nociceptor neurons affect cancer immunosurveillance. Nature. 2022;611:405–12.
- 20. Magnon C, Hondermarck H. The neural addiction of cancer. Nat Rev Cancer. 2023;23:317–34.
- Tibensky M, Mravec B. Role of the parasympathetic nervous system in cancer initiation and progression. Clin Transl Oncol. 2021;23:669–81.
- 22. Chen H, Wang C, Chen Z, Huang T, Lin Y, Chen J, Zhang B, He X. The depth of perineural invasion is an independent prognostic factor for stage II colorectal cancer. BMC Cancer. 2024;24:433.
- Wang H, Zheng Q, Lu Z, Wang L, Ding L, Xia L, Zhang H, Wang M, Chen Y, Li G. Role of the nervous system in cancers: a review. Cell Death Discov. 2021;7:76.
- Fjæstad KY, Rømer AMA, Goitea V, Johansen AZ, Thorseth ML, Carretta M, Engelholm LH, Grøntved L, Junker N, Madsen DH. Blockade of beta-adrenergic receptors reduces cancer growth and enhances the response to anti-CTLA4 therapy by modulating the tumor microenvironment. Oncogene. 2022;41:1364–75.
- 25. Passang T, Wang S, Zhang H, Zeng F, Hsu PC, Wang W, Li JM, Liu Y, Ravindranathan S, Lesinski GB, Waller EK: VPAC2 Receptor Signaling Promotes Growth and Immunosuppression in Pancreatic Cancer. Cancer Res 2024, null.

- Allen JK, Armaiz-Pena GN, Nagaraja AS, Sadaoui NC, Ortiz T, Dood R, Ozcan M, Herder DM, Haemmerle M, Gharpure KM, et al. Sustained Adrenergic Signaling Promotes Intratumoral Innervation through BDNF Induction. Cancer Res. 2018;78:3233–42.
- Renz BW, Takahashi R, Tanaka T, Macchini M, Hayakawa Y, Dantes Z, Maurer HC, Chen X, Jiang Z, Westphalen CB, et al. β2 Adrenergic-Neurotrophin Feedforward Loop Promotes Pancreatic Cancer. Cancer Cell. 2018;33:75-90.e77.
- Lawn S, Krishna N, Pisklakova A, Qu X, Fenstermacher DA, Fournier M, Vrionis FD, Tran N, Chan JA, Kenchappa RS, Forsyth PA. Neurotrophin signaling via TrkB and TrkC receptors promotes the growth of brain tumor-initiating cells. J Biol Chem. 2015;290:3814–24.
- 29. Wang Z, Liu W, Wang C, Li Y, Ai Z. Acetylcholine promotes the selfrenewal and immune escape of CD133+ thyroid cancer cells through activation of CD133-Akt pathway. Cancer Lett. 2020;471:116–24.
- 30. Huang D, Wang Y, Thompson JW, Yin T, Alexander PB, Qin D, Mudgal P, Wu H, Liang Y, Tan L, et al. Cancer-cell-derived GABA promotes β -catenin-mediated tumour growth and immunosuppression. Nat Cell Biol. 2022;24:230–41.
- Guo X, Pan Y, Xiong M, Sanapala S, Anastasaki C, Cobb O, Dahiya S, Gutmann DH. Midkine activation of CD8(+) T cells establishes a neuron-immune-cancer axis responsible for low-grade glioma growth. Nat Commun. 2020;11:2177.
- Zahalka AH, Arnal-Estapé A, Maryanovich M, Nakahara F, Cruz CD, Finley LWS, Frenette PS. Adrenergic nerves activate an angio-metabolic switch in prostate cancer. Science. 2017;358:321–6.
- S S, A B, B L: Involvement of neuronal factors in tumor angiogenesis and the shaping of the cancer microenvironment. Frontiers in immunology 2024, 15:1284629.
- 34. Guan F, Wu X, Zhou J, Lin Y, He Y, Fan C, Zeng Z, Xiong W. Mitochondrial transfer in tunneling nanotubes-a new target for cancer therapy. J Exp Clin Cancer Res. 2024;43:147.
- Wang F, Chen X, Cheng H, Song L, Liu J, Caplan S, Zhu L, Wu JY. MICAL2PV suppresses the formation of tunneling nanotubes and modulates mitochondrial trafficking. EMBO Rep. 2021;22: e52006.
- Jung E, Osswald M, Blaes J, Wiestler B, Sahm F, Schmenger T, Solecki G, Deumelandt K, Kurz FT, Xie R, et al. Tweety-Homolog 1 Drives Brain Colonization of Gliomas. J Neurosci. 2017;37:6837–50.
- Deborde S, Gusain L, Powers A, Marcadis A, Yu Y, Chen CH, Frants A, Kao E, Tang LH, Vakiani E, et al. Reprogrammed Schwann Cells Organize into Dynamic Tracks that Promote Pancreatic Cancer Invasion. Cancer Discov. 2022;12:2454–73.
- He K, Wang H, Huo R, Jiang SH, Xue J. Schwann cells and enteric glial cells: Emerging stars in colorectal cancer. Biochim Biophys Acta Rev Cancer. 2024;1879: 189160.
- Su D, Guo X, Huang L, Ye H, Li Z, Lin L, Chen R, Zhou Q. Tumor-neuroglia interaction promotes pancreatic cancer metastasis. Theranostics. 2020;10:5029–47.
- Hsu YL, Hou MF, Kuo PL, Huang YF, Tsai EM. Breast tumor-associated osteoblast-derived CXCL5 increases cancer progression by ERK/MSK1/ Elk-1/snail signaling pathway. Oncogene. 2013;32:4436–47.
- Zhi X, Wu F, Qian J, Ochiai Y, Lian G, Malagola E, Zheng B, Tu R, Zeng Y, Kobayashi H, et al: Nociceptive neurons promote gastric tumour progression via a CGRP-RAMP1 axis. Nature 2025.
- 42. Lei Y, He X, Huang H, He Y, Lan J, Yang J, Liu W, Zhang T. Nerve growth factor orchestrates NGAL and matrix metalloproteinases activity to promote colorectal cancer metastasis. Clin Transl Oncol. 2022;24:34–47.
- Fu Y, Shen K, Wang H, Wang S, Wang X, Zhu L, Zheng Y, Zou T, Ci H, Dong Q, Qin LX. Alpha5 nicotine acetylcholine receptor subunit promotes intrahepatic cholangiocarcinoma metastasis. Signal Transduct Target Ther. 2024;9:63.
- Xu Q, Wang Z, Chen X, Duan W, Lei J, Zong L, Li X, Sheng L, Ma J, Han L, et al. Stromal-derived factor-1α/CXCL12-CXCR4 chemotactic pathway promotes perineural invasion in pancreatic cancer. Oncotarget. 2015;6:4717–32.
- Zaitseva L, Murray MY, Shafat MS, Lawes MJ, MacEwan DJ, Bowles KM, Rushworth SA. Ibrutinib inhibits SDF1/CXCR4 mediated migration in AML. Oncotarget. 2014;5:9930–8.
- Tian Z, Ou G, Su M, Li R, Pan L, Lin X, Zou J, Chen S, Li Y, Huang K, Chen Y. TIMP1 derived from pancreatic cancer cells stimulates Schwann cells and promotes the occurrence of perineural invasion. Cancer Lett. 2022;546: 215863.

- Guo K, Ma Q, Li J, Wang Z, Shan T, Li W, Xu Q, Xie K. Interaction of the sympathetic nerve with pancreatic cancer cells promotes perineural invasion through the activation of STAT3 signaling. Mol Cancer Ther. 2013;12:264–73.
- NS J, JT D, PK M, N F, D Y, A P, AF Z, D V, KQ T, M Z, et al: A drug repositioning approach identifies tricyclic antidepressants as inhibitors of small cell lung cancer and other neuroendocrine tumors. Cancer discovery 2013, 3:1364–1377.
- Blackhall F, Jao K, Greillier L, Cho BC, Penkov K, Reguart N, Majem M, Nackaerts K, Syrigos K, Hansen K, et al. Efficacy and Safety of Rovalpituzumab Tesirine Compared With Topotecan as Second-Line Therapy in DLL3-High SCLC: Results From the Phase 3 TAHOE Study. J Thorac Oncol. 2021;16:1547–58.
- Fabrizio FP, Sparaneo A, Gorgoglione G, Battista P, Centra F, Delli Muti F, Trombetta D, Centonza A, Graziano P, Rossi A, et al: Effects of KEAP1 Silencing on NRF2 and NOTCH Pathways in SCLC Cell Lines. Cancers (Basel) 2024, 16.
- H X, G C, K F, W G, F Q: Neuronatin Promotes the Progression of Nonsmall Cell Lung Cancer by Activating the NF-kB Signaling. Current cancer drug targets 2024, 24:1128–1143.
- Mc Z. P X, GL L, M Z: Could lung cancer exosomes induce apoptosis of natural killer cells through the p75NTR-proNGF-sortilin axis? Med Hypotheses. 2017;108:151–3.
- F G, N G, S F, CW R, L W, S R, RF T, A F, P J, MM W, H H: The neurotrophic tyrosine kinase receptor TrkA and its ligand NGF are increased in squamous cell carcinomas of the lung. Scientific reports 2018, 8:8135.
- 54. Chen WY, Wen YC, Lin SR, Yeh HL, Jiang KC, Chen WH, Lin YS, Zhang Q, Liew PL, Hsiao M, et al. Nerve growth factor interacts with CHRM4 and promotes neuroendocrine differentiation of prostate cancer and castration resistance. Commun Biol. 2021;4:22.
- L X, S L, Y L, Y H, B N, L W, H M, X L, Z C, Z L: NDNF inhibits the migration and invasion of human renal cancer cells through epithelial-mesenchymal transition. Oncology letters 2019, 17:2969–2975.
- M J, Y W, T Z, W L, Q W: Norepinephrine/β 2 -Adrenergic Receptor Pathway Promotes the Cell Proliferation and Nerve Growth Factor Production in Triple-Negative Breast Cancer. Journal of breast cancer 2023, 26:268–285.
- J C, S T, M P, G T, L D, E H, N B, A VO, L D, RP B, et al: ProNGF promotes brain metastasis through TrkA/EphA2 induced Src activation in triple negative breast cancer cells. Experimental hematology & oncology 2023, 12:104.
- Q C, D J, Y Z, C C: The tumor-nerve circuit in breast cancer. Cancer metastasis reviews 2023, 42:543–574.
- D Z, X L, Z Y, C W, N N, J L: GABAergic signaling facilitates breast cancer metastasis by promoting ERK1/2-dependent phosphorylation. Cancer letters 2014, 348:100–108.
- 60. Ferraguti G, Terracina S, Tarani L, Fanfarillo F, Allushi S, Caronti B, Tirassa P, Polimeni A, Lucarelli M, Cavalcanti L, et al. Nerve Growth Factor and the Role of Inflammation in Tumor Development. Curr Issues Mol Biol. 2024;46:965–89.
- Benzaquen D, Lawrence YR, Taussky D, Zwahlen D, Oehler C, Champion A: The Crosstalk between Nerves and Cancer-A Poorly Understood Phenomenon and New Possibilities. Cancers (Basel) 2024, 16.
- Bruno F, Arcuri D, Vozzo F, Malvaso A, Montesanto A, Maletta R. Expression and Signaling Pathways of Nerve Growth Factor (NGF) and Pro-NGF in Breast Cancer: A Systematic Review. Curr Oncol. 2022;29:8103–20.
- 63. Han S, Wang D, Huang Y, Zeng Z, Xu P, Xiong H, Ke Z, Zhang Y, Hu Y, Wang F, et al. A reciprocal feedback between colon cancer cells and Schwann cells promotes the proliferation and metastasis of colon cancer. J Exp Clin Cancer Res. 2022;41:348.
- 64. Pundavela J, Demont Y, Jobling P, Lincz LF, Roselli S, Thorne RF, Bond D, Bradshaw RA, Walker MM, Hondermarck H. ProNGF correlates with Gleason score and is a potential driver of nerve infiltration in prostate cancer. Am J Pathol. 2014;184:3156–62.
- 65. Lai PC, Chiu TH, Huang YT. Overexpression of BDNF and TrkB in human bladder cancer specimens. Oncol Rep. 2010;24:1265–70.
- Çerçi B, Gök A, Akyol A. Brain-derived neurotrophic factor: Its role in energy balance and cancer cachexia. Cytokine Growth Factor Rev. 2023;71–72:105–16.

- Tajbakhsh A, Mokhtari-Zaer A, Rezaee M, Afzaljavan F, Rivandi M, Hassanian SM, Ferns GA, Pasdar A, Avan A. Therapeutic Potentials of BDNF/ TrkB in Breast Cancer; Current Status and Perspectives. J Cell Biochem. 2017;118:2502–15.
- Dun XP, Parkinson DB. Classic axon guidance molecules control correct nerve bridge tissue formation and precise axon regeneration. Neural Regen Res. 2020;15:6–9.
- 69. Mancino M, Ametller E, Gascón P, Almendro V. The neuronal influence on tumor progression. Biochim Biophys Acta. 2011;1816:105–18.
- Arese M, Bussolino F, Pergolizzi M, Bizzozero L, Pascal D. Tumor progression: the neuronal input. Ann Transl Med. 2018;6:89.
- Ayala GE, Dai H, Powell M, Li R, Ding Y, Wheeler TM, Shine D, Kadmon D, Thompson T, Miles BJ, et al. Cancer-related axonogenesis and neurogenesis in prostate cancer. Clin Cancer Res. 2008;14:7593–603.
- Madeo M, Colbert PL, Vermeer DW, Lucido CT, Cain JT, Vichaya EG, Grossberg AJ, Muirhead D, Rickel AP, Hong Z, et al. Cancer exosomes induce tumor innervation. Nat Commun. 2018;9:4284.
- Moore SW, Tessier-Lavigne M, Kennedy TE. Netrins and their receptors. Adv Exp Med Biol. 2007;621:17–31.
- Webber CA, Christie KJ, Cheng C, Martinez JA, Singh B, Singh V, Thomas D, Zochodne DW. Schwann cells direct peripheral nerve regeneration through the Netrin-1 receptors, DCC and Unc5H2. Glia. 2011;59:1503–17.
- Lu R, Fan C, Shangguan W, Liu Y, Li Y, Shang Y, Yin D, Zhang S, Huang Q, Li X, et al. Neurons generated from carcinoma stem cells support cancer progression. Signal Transduct Target Ther. 2017;2:16036.
- Demir E, Schorn S, Schremmer-Danninger E, Wang K, Kehl T, Giese NA, Algül H, Friess H, Ceyhan GO. Perineural mast cells are specifically enriched in pancreatic neuritis and neuropathic pain in pancreatic cancer and chronic pancreatitis. PLoS ONE. 2013;8: e60529.
- Tang PC, Chung JY, Liao J, Chan MK, Chan AS, Cheng G, Li C, Huang XR, Ng CS, Lam EW, et al: Single-cell RNA sequencing uncovers a neuronlike macrophage subset associated with cancer pain. Sci Adv 2022, 8:eabn5535.
- Ma H, Pan Z, Lai B, Li M, Wang J. Contribution of immune cells to cancer-related neuropathic pain: An updated review. Mol Pain. 2023;19:17448069231182236.
- 79. Sánchez ML, Rodríguez FD, Coveñas R: Neuropeptide Y Peptide Family and Cancer: Antitumor Therapeutic Strategies. Int J Mol Sci 2023, 24.
- Diaz-delCastillo M, Christiansen SH, Appel CK, Falk S, Woldbye DPD, Heegaard AM. Neuropeptide Y is Up-regulated and Induces Antinociception in Cancer-induced Bone Pain. Neuroscience. 2018;384:111–9.
- 81. Ye Y, Xie T, Amit M. Targeting the Nerve-Cancer Circuit. Cancer Res. 2023;83:2445–7.
- Yang Y, Yang W, Zhang R, Wang Y. Peripheral Mechanism of Cancer-Induced Bone Pain. Neurosci Bull. 2024;40:815–30.
- Alrawashdeh W, Jones R, Dumartin L, Radon TP, Cutillas PR, Feakins RM, Dmitrovic B, Demir IE, Ceyhan GO, Crnogorac-Jurcevic T. Perineural invasion in pancreatic cancer: proteomic analysis and in vitro modelling. Mol Oncol. 2019;13:1075–91.
- 84. Yoneda T, Hiasa M, Okui T. Crosstalk Between Sensory Nerves and Cancer in Bone. Curr Osteoporos Rep. 2018;16:648–56.
- Lozano-Ondoua AN, Symons-Liguori AM, Vanderah TW: Cancerinduced bone pain: Mechanisms and models. Neurosci Lett 2013, 557 Pt A:52–59.
- Sevcik MA, Ghilardi JR, Peters CM, Lindsay TH, Halvorson KG, Jonas BM, Kubota K, Kuskowski MA, Boustany L, Shelton DL, Mantyh PW. Anti-NGF therapy profoundly reduces bone cancer pain and the accompanying increase in markers of peripheral and central sensitization. Pain. 2005;115:128–41.
- Barnes PJ, Belvisi MG, Rogers DF. Modulation of neurogenic inflammation: novel approaches to inflammatory disease. Trends Pharmacol Sci. 1990;11:185–9.
- Cruceriu D, Baldasici O, Balacescu O, Berindan-Neagoe I. The dual role of tumor necrosis factor-alpha (TNF-a) in breast cancer: molecular insights and therapeutic approaches. Cell Oncol (Dordr). 2020;43:1–18.
- 89. Caronni N, La Terza F, Vittoria FM, Barbiera G, Mezzanzanica L, Cuzzola V, Barresi S, Pellegatta M, Canevazzi P, Dunsmore G, et al. IL-1 β (+) macrophages fuel pathogenic inflammation in pancreatic cancer. Nature. 2023;623:415–22.

- 91. Erin N, Szallasi A: Carcinogenesis and Metastasis: Focus on TRPV1-Positive Neurons and Immune Cells. Biomolecules 2023, 13.
- Bujak JK, Kosmala D, Szopa IM, Majchrzak K, Bednarczyk P. Inflammation, Cancer and Immunity-Implication of TRPV1 Channel. Front Oncol. 2019;9:1087.
- Jiang M, Wang Y, Wang J, Feng S, Wang X. The etiological roles of miRNAs, IncRNAs, and circRNAs in neuropathic pain: A narrative review. J Clin Lab Anal. 2022;36: e24592.
- 94. Sakai A, Suzuki H. microRNA and Pain. Adv Exp Med Biol. 2015;888:17–39.
- Pontecorvi G, Bellenghi M, Puglisi R, Carè A, Mattia G. Tumor-derived extracellular vesicles and microRNAs: Functional roles, diagnostic, prognostic and therapeutic options. Cytokine Growth Factor Rev. 2020;51:75–83.
- Ruan X, Yan W, Cao M, Daza RAM, Fong MY, Yang K, Wu J, Liu X, Palomares M, Wu X, et al. Breast cancer cell-secreted miR-199b-5p hijacks neurometabolic coupling to promote brain metastasis. Nat Commun. 2024;15:4549.
- Elramah S, López-González MJ, Bastide M, Dixmérias F, Roca-Lapirot O, Wielanek-Bachelet AC, Vital A, Leste-Lasserre T, Brochard A, Landry M, Favereaux A. Spinal miRNA-124 regulates synaptopodin and nociception in an animal model of bone cancer pain. Sci Rep. 2017;7:10949.
- Bali KK, Selvaraj D, Satagopam VP, Lu J, Schneider R, Kuner R. Genomewide identification and functional analyses of microRNA signatures associated with cancer pain. EMBO Mol Med. 2013;5:1740–58.
- Amit M, Takahashi H, Dragomir MP, Lindemann A, Gleber-Netto FO, Pickering CR, Anfossi S, Osman AA, Cai Y, Wang R, et al. Loss of p53 drives neuron reprogramming in head and neck cancer. Nature. 2020;578:449–54.
- Renz BW, Tanaka T, Sunagawa M, Takahashi R, Jiang Z, Macchini M, Dantes Z, Valenti G, White RA, Middelhoff MA, et al. Cholinergic Signaling via Muscarinic Receptors Directly and Indirectly Suppresses Pancreatic Tumorigenesis and Cancer Stemness. Cancer Discov. 2018;8:1458–73.
- 101. Yu H, Xia H, Tang Q, Xu H, Wei G, Chen Y, Dai X, Gong Q, Bi F. Acetylcholine acts through M3 muscarinic receptor to activate the EGFR signaling and promotes gastric cancer cell proliferation. Sci Rep. 2017;7:40802.
- 102. Kniewallner KM, Grimm N, Humpel C. Platelet-derived nerve growth factor supports the survival of cholinergic neurons in organotypic rat brain slices. Neurosci Lett. 2014;574:64–9.
- Sampaio Moura N, Schledwitz A, Alizadeh M, Kodan A, Njei LP, Raufman JP: Cholinergic Mechanisms in Gastrointestinal Neoplasia. Int J Mol Sci 2024, 25.
- Al-Wadei MH, Banerjee J, Al-Wadei HA, Schuller HM. Nicotine induces self-renewal of pancreatic cancer stem cells via neurotransmitter-driven activation of sonic hedgehog signalling. Eur J Cancer. 2016;52:188–96.
- Al-Wadei MH, Al-Wadei HA, Schuller HM. Effects of chronic nicotine on the autocrine regulation of pancreatic cancer cells and pancreatic duct epithelial cells by stimulatory and inhibitory neurotransmitters. Carcinogenesis. 2012;33:1745–53.
- Ascano M Jr, Mukherjee N, Bandaru P, Miller JB, Nusbaum JD, Corcoran DL, Langlois C, Munschauer M, Dewell S, Hafner M, et al. FMRP targets distinct mRNA sequence elements to regulate protein expression. Nature. 2012;492:382–6.
- 107. Y H, MR B, YT, S M, GB G, KM C, KM R-L, ML K, J B-C, CM T, et al: Platelet-Derived Growth Factor Receptor-α Regulates Proliferation of Gastrointestinal Stromal Tumor Cells With Mutations in KIT by Stabilizing ETV1. 2015, 149:420–432.e416.
- WK K, M P, YK K, YK T, HK Y, JM L, Research KHJCcraojotAAfC: Micro-RNA-494 downregulates KIT and inhibits gastrointestinal stromal tumor cell proliferation. 2011, 17:7584–7594.
- 109. Yonekura S, Terrisse S, Alves Costa Silva C, Lafarge A, lebba V, Ferrere G, Goubet AG, Fahrner JE, Lahmar I, Ueda K, et al: Cancer induces a stress ileopathy depending on B-adrenergic receptors and promoting dysbiosis that contribute to carcinogenesis. Cancer Discov 2021.
- 110. Zong C, Yang M, Guo X, Ji W. Chronic restraint stress promotes gastric epithelial malignant transformation by activating the Akt/p53 signaling pathway via ADRB2. Oncol Lett. 2022;24:300.

- 111. Feng J, Zhang Y, She X, Sun Y, Fan L, Ren X, Fu H, Liu C, Li P, Zhao C, et al. Hypermethylated gene ANKDD1A is a candidate tumor suppressor that interacts with FIH1 and decreases HIF1α stability to inhibit cell autophagy in the glioblastoma multiforme hypoxia microenvironment. Oncogene. 2019;38:103–19.
- 112. Wu FQ, Fang T, Yu LX, Lv GS, Lv HW, Liang D, Li T, Wang CZ, Tan YX, Ding J, et al. ADRB2 signaling promotes HCC progression and sorafenib resistance by inhibiting autophagic degradation of HIF1a. J Hepatol. 2016;65:314–24.
- 113. Zhou Z, Zhan J, Luo Q, Hou X, Wang S, Xiao D, Xie Z, Liang H, Lin S, Zheng M. ADRB3 induces mobilization and inhibits differentiation of both breast cancer cells and myeloid-derived suppressor cells. Cell Death Dis. 2022;13:141.
- 114. R G, Z J: Genetic manipulations of AMPA glutamate receptors in hippocampal synaptic plasticity. Neuropharmacology 2021, 194:108630.
- L J, N L, F Z, B H, D K, P Z, X L: Discovery of GluN2A subtype-selective N -methyl-d-aspartate (NMDA) receptor ligands. Acta pharmaceutica Sinica B 2024, 14:1987–2005.
- CAM dL-L, M C-R, ZU K: AMPA Receptors in Synaptic Plasticity, Memory Function, and Brain Diseases. Cellular and molecular neurobiology 2025, 45:14.
- 117. Li F, He C, Yao H, Zhao Y, Ye X, Zhou S, Zou J, Li Y, Li J, Chen S, et al. Glutamate from nerve cells promotes perineural invasion in pancreatic cancer by regulating tumor glycolysis through HK2 mRNA-m6A modification. Pharmacol Res. 2023;187: 106555.
- 118. Li L, Hanahan D. Hijacking the neuronal NMDAR signaling circuit to promote tumor growth and invasion. Cell. 2013;153:86–100.
- 119. Li L, Zeng Q, Bhutkar A, Galván JA, Karamitopoulou E, Noordermeer D, Peng MW, Piersigilli A, Perren A, Zlobec I, et al. GKAP Acts as a Genetic Modulator of NMDAR Signaling to Govern Invasive Tumor Growth. Cancer Cell. 2018;33:736-751.e735.
- Herner A, Sauliunaite D, Michalski CW, Erkan M, De Oliveira T, Abiatari I, Kong B, Esposito I, Friess H, Kleeff J. Glutamate increases pancreatic cancer cell invasion and migration via AMPA receptor activation and Kras-MAPK signaling. Int J Cancer. 2011;129:2349–59.
- 121. Zheng M, Zhou Z, Tian X, Xiao D, Hou X, Xie Z, Liang H, Lin S. ADRB3 expression in tumor cells is a poor prognostic factor and promotes proliferation in non-small cell lung carcinoma. Cancer Immunol Immunother. 2020;69:2345–55.
- Zhao S, Shi J, Yu G, Li D, Wang M, Yuan C, Zhou H, Parizadeh A, Li Z, Guan MX, Ye S. Photosensitive tyrosine analogues unravel site-dependent phosphorylation in TrkA initiated MAPK/ERK signaling. Commun Biol. 2020;3:706.
- 123. TI O, YM L, TJ N, YS K, S M, J K, Y K, RH R, YJ K, S H, JH L: Fascaplysin Exerts Anti-Cancer Effects through the Downregulation of Survivin and HIF-1α and Inhibition of VEGFR2 and TRKA. International journal of molecular sciences 2017, 18.
- 124. Ef W. AR N: Signal integration by JNK and p38 MAPK pathways in cancer development. Nat Rev Cancer. 2009;9:537–49.
- 125. M W, RJ D: The native structure of the activated Raf protein kinase is a membrane-bound multi-subunit complex. The Journal of biological chemistry 1994, 269:6695–6701.
- 126. AS A: PI3K/Akt/mTOR inhibitors in cancer: At the bench and bedside. Seminars in cancer biology 2019, 59:125–132.
- 127. Mk E, Kh T. SR S: Role of the PI3K/AKT/mTOR signaling pathway in ovarian cancer: Biological and therapeutic significance. Semin Cancer Biol. 2019;59:147–60.
- Hj J, Pg S, Yj L, Kj S. L C, YC C: PLCγ1: Potential arbitrator of cancer progression. Advances in biological regulation. 2018;67:179–89.
- W B, HH W, FJ T, XY H, MT Q, JY W, HJ Z, LH W, XP W: A TrkB-STAT3-miR-204–5p regulatory circuitry controls proliferation and invasion of endometrial carcinoma cells. Molecular cancer 2013, 12:155.
- 130. M M, SS N, E S, M M, A S, N R: BDNF and its signaling in cancer. Journal of cancer research and clinical oncology 2023, 149:2621–2636.
- D S, O B, H P, TA O, MT T, AC J, DE C: S100A16 promotes differentiation and contributes to a less aggressive tumor phenotype in oral squamous cell carcinoma. BMC cancer 2015, 15:631.
- MA DIC-M, J B, R S-P, S S, T N, A G, A P, P S, A L, A D, et al: p75 neurotrophin receptor and pro-BDNF promote cell survival and migration in clear cell renal cell carcinoma. Oncotarget 2016, 7:34480–34497.

- CA B, NM M, K M, E W, BD E, BR C: Rescue of NMDAR-dependent synaptic plasticity in Fmr1 knock-out mice. Cerebral cortex (New York, NY : 1991) 2015, 25:271–279.
- Lee KY, Wang H, Yook Y, Rhodes JS, Christian-Hinman CA, Tsai NP. Tumor suppressor p53 modulates activity-dependent synapse strengthening, autism-like behavior and hippocampus-dependent learning. Mol Psychiatry. 2023;28:3782–94.
- Piao Y, Lu L, de Groot J. AMPA receptors promote perivascular glioma invasion via beta1 integrin-dependent adhesion to the extracellular matrix. Neuro Oncol. 2009;11:260–73.
- Smith CS, Álvarez Z, Qiu R, Sasselli IR, Clemons T, Ortega JA, Vilela-Picos M, Wellman H, Kiskinis E, Stupp SI. Enhanced Neuron Growth and Electrical Activity by a Supramolecular Netrin-1 Mimetic Nanofiber. ACS Nano. 2023;17:19887–902.
- 137. Kryza D, Wischhusen J, Richaud M, Hervieu M, Sidi Boumedine J, Delcros JG, Besse S, Baudier T, Laval PA, Breusa S, et al. From netrin-1-targeted SPECT/CT to internal radiotherapy for management of advanced solid tumors. EMBO Mol Med. 2023;15: e16732.
- 138. Ye X, Qiu Y, Gao Y, Wan D, Zhu H. A Subtle Network Mediating Axon Guidance: Intrinsic Dynamic Structure of Growth Cone, Attractive and Repulsive Molecular Cues, and the Intermediate Role of Signaling Pathways. Neural Plast. 2019;2019:1719829.
- Gong Z, Zhang Y, Wang W, Li X, Wang K, You X, Wu J. Netrin-1 Role in Nociceptive Neuron Sprouting through Activation of DCC Signaling in a Rat Model of Bone Cancer Pain. J Integr Neurosci. 2024;23:47.
- Furne C, Rama N, Corset V, Chédotal A, Mehlen P. Netrin-1 is a survival factor during commissural neuron navigation. Proc Natl Acad Sci U S A. 2008;105:14465–70.
- 141. Huang Y, Zhou X, Liu J, Cao Y, Fu W, Yang J. Emerging neuroimmune mechanisms in cancer neuroscience. Cancer Lett. 2025;612: 217492.
- 142. Zhang Y, Liao Q, Wen X, Fan J, Yuan T, Tong X, Jia R, Chai P, Fan X. Hijacking of the nervous system in cancer: mechanism and therapeutic targets. Mol Cancer. 2025;24:44.
- 143. Qin JF, Jin FJ, Li N, Guan HT, Lan L, Ni H, Wang Y. Adrenergic receptor β2 activation by stress promotes breast cancer progression through macrophages M2 polarization in tumor microenvironment. BMB Rep. 2015;48:295–300.
- 144. Xia Y, Wei Y, Li ZY, Cai XY, Zhang LL, Dong XR, Zhang S, Zhang RG, Meng R, Zhu F, Wu G. Catecholamines contribute to the neovascularization of lung cancer via tumor-associated macrophages. Brain Behav Immun. 2019;81:111–21.
- Martinelli S, Amore F, Canu L, Maggi M, Rapizzi E. Tumour microenvironment in pheochromocytoma and paraganglioma. Front Endocrinol (Lausanne). 2023;14:1137456.
- 146. Zhang B, Guo X, Huang L, Zhang Y, Li Z, Su D, Lin L, Zhou P, Ye H, Lu Y, Zhou Q. Tumour-associated macrophages and Schwann cells promote perineural invasion via paracrine loop in pancreatic ductal adenocarcinoma. Br J Cancer. 2024;130:542–54.
- Nishikawa H, Koyama S: Mechanisms of regulatory T cell infiltration in tumors: implications for innovative immune precision therapies. J Immunother Cancer 2021, 9.
- Shackleton EG, Ali HY, Khan M, Pockley GA, McArdle SE: Novel Combinatorial Approaches to Tackle the Immunosuppressive Microenvironment of Prostate Cancer. Cancers (Basel) 2021, 13.
- 149. Verona F, Di Bella S, Schirano R, Manfredi C, Angeloro F, Bozzari G, Todaro M, Giannini G, Stassi G, Veschi V. Cancer stem cells and tumorassociated macrophages as mates in tumor progression: mechanisms of crosstalk and advanced bioinformatic tools to dissect their phenotypes and interaction. Front Immunol. 2025;16:1529847.
- Lin S, Liu D, Liang T, Zhuang Y, Wang X, Ma S, Li Q, Hu K. Cryoablationinduced modulation of Treg cells and the TGF-β pathway in lung adenocarcinoma: implications for increased antitumor immunity. BMC Med. 2025;23:89.
- Cervantes-Villagrana RD, Albores-García D, Cervantes-Villagrana AR, García-Acevez SJ. Tumor-induced neurogenesis and immune evasion as targets of innovative anti-cancer therapies. Signal Transduct Target Ther. 2020;5:99.
- 152. Wu Y, Ma J, Yang X, Nan F, Zhang T, Ji S, Rao D, Feng H, Gao K, Gu X, et al. Neutrophil profiling illuminates anti-tumor antigen-presenting potency. Cell. 2024;187:1422-1439.e1424.

- Zhong J, Xing X, Gao Y, Pei L, Lu C, Sun H, Lai Y, Du K, Xiao F, Yang Y, et al. Distinct roles of TREM2 in central nervous system cancers and peripheral cancers. Cancer Cell. 2024;42:968-984.e969.
- Jiang L, Cai S, Weng Z, Zhang S, Jiang SH. Peripheral, central, and chemotherapy-induced neuropathic changes in pancreatic cancer. Trends Neurosci. 2025;48:124–39.
- 155. Zhang L, Pan J, Wang M, Yang J, Zhu S, Li L, Hu X, Wang Z, Pang L, Li P, et al. Chronic Stress-Induced and Tumor Derived SP1(+) Exosomes Polarizing IL-1 β (+) Neutrophils to Increase Lung Metastasis of Breast Cancer. Adv Sci (Weinh). 2025;12: e2310266.
- 156. Pan J, Zhang L, Wang X, Li L, Yang C, Wang Z, Su K, Hu X, Zhang Y, Ren G, et al. Chronic stress induces pulmonary epithelial cells to produce acetylcholine that remodels lung pre-metastatic niche of breast cancer by enhancing NETosis. J Exp Clin Cancer Res. 2023;42:255.
- 157. Tabei Y, Nakajima Y. IL-1β-activated PI3K/AKT and MEK/ERK pathways coordinately promote induction of partial epithelial-mesenchymal transition. Cell Commun Signal. 2024;22:392.
- 158. Brain SD, Grant AD. Vascular actions of calcitonin gene-related peptide and adrenomedullin. Physiol Rev. 2004;84:903–34.
- Waldmann TA. The biology of IL-15: implications for cancer therapy and the treatment of autoimmune disorders. J Investig Dermatol Symp Proc. 2013;16:S28-30.
- 160. Wang K, Ni B, Xie Y, Li Z, Yuan L, Meng C, Zhao T, Gao S, Huang C, Wang H, et al: Nociceptor neurons promote PDAC progression and cancer pain by interaction with cancer-associated fibroblasts and suppression of natural killer cells. Cell Res 2025.
- Oya Y, Hayakawa Y, Koike K. Tumor microenvironment in gastric cancers. Cancer Sci. 2020;111:2696–707.
- 162. Yu Z, Li H, Wang M, Luo W, Xue Y: GDNF regulates lipid metabolism and glioma growth through RET/ERK/HIF-1/SREBP-1. Int J Oncol 2022, 61.
- Li Y, Zhang C, Jiang A, Lin A, Liu Z, Cheng X, Wang W, Cheng Q, Zhang J, Wei T, Luo P. Potential anti-tumor effects of regulatory T cells in the tumor microenvironment: a review. J Transl Med. 2024;22:293.
- 164. Xie SZ, Yang LY, Wei R, Shen XT, Pan JJ, Yu SZ, Zhang C, Xu H, Xu JF, Zheng X, et al. Targeting SPP1-orchestrated neutrophil extracellular traps-dominant pre-metastatic niche reduced HCC lung metastasis. Exp Hematol Oncol. 2024;13:111.
- 165. Wu Z, Shan Q, Jiang Y, Huang W, Wang Z, Zhuang Y, Liu J, Li T, Yang Z, Li C, et al. Irreversible electroporation combined with PD-L1/IL-6 dual blockade promotes anti-tumor immunity via cDC2/CD4(+)T cell axis in MHC-I deficient pancreatic cancer. Cancer Lett. 2025;617: 217620.
- 166. SO M, Toxins JBJ: Botulinum Neurotoxins and Cancer-A Review of the Literature. 2020, 12.
- JL T, R S, S LC, SC D, MS M, S K, S P, S G, KR R, KM H, et al: Metabotropic glutamate receptor 1 disrupts mammary acinar architecture and initiates malignant transformation of mammary epithelial cells. 2015, 151:57–73.
- 168. Q C, D J, Y Z, reviews CCJCm: The tumor-nerve circuit in breast cancer. 2023, 42:543–574.
- Kojadinovic A, Laderian B, Mundi PS. Targeting TRK: A fast-tracked application of precision oncology and future directions. Crit Rev Oncol Hematol. 2021;165: 103451.
- Pan Y, Hysinger JD, Barron T, Schindler NF, Cobb O, Guo X, Yalçın B, Anastasaki C, Mulinyawe SB, Ponnuswami A, et al. NF1 mutation drives neuronal activity-dependent initiation of optic glioma. Nature. 2021;594:277–82.
- 171. Liu Q, Tan J, Zhao Z, Li R, Zheng L, Chen X, Li L, Dong X, Wen T, Liu J: Combined Usage of MDK Inhibitor Augments Interferon-γ Anti-Tumor Activity in the SKOV3 Human Ovarian Cancer Cell Line. Biomedicines 2022, 11.
- Flügel A, Schwaiger FW, Neumann H, Medana I, Willem M, Wekerle H, Kreutzberg GW, Graeber MB. Neuronal FasL induces cell death of encephalitogenic T lymphocytes. Brain Pathol. 2000;10:353–64.
- Upadhyay R, Boiarsky JA, Pantsulaia G, Svensson-Arvelund J, Lin MJ, Wroblewska A, Bhalla S, Scholler N, Bot A, Rossi JM, et al. A Critical Role for Fas-Mediated Off-Target Tumor Killing in T-cell Immunotherapy. Cancer Discov. 2021;11:599–613.
- 174. Liu D, Vadgama J, Wu Y. Basal-like breast cancer with low TGF β and high TNF α pathway activity is rich in activated memory CD4 T cells and has a good prognosis. Int J Biol Sci. 2021;17:670–82.

- 175. Shan ZG, Zhao YL, Zhang JY, Yan ZB, Wang TT, Mao FY, Teng YS, Peng LS, Chen WY, Wang P, et al. FasL(+) PD-L2(+) Identifies a Novel Immunosuppressive Neutrophil Population in Human Gastric Cancer That Promotes Disease Progression. Adv Sci (Weinh). 2022;9: e2103543.
- Sun X, Lin M, Tian Z, Ma Y, Lv L. GABA/baclofen stabilizes PD-L1 and enhances immunotherapy of breast cancer. Heliyon. 2024;10: e28600.
- 177. Murugan S, Rousseau B, Sarkar DK: Beta 2 Adrenergic Receptor Antagonist Propranolol and Opioidergic Receptor Antagonist Naltrexone Produce Synergistic Effects on Breast Cancer Growth Prevention by Acting on Cancer Cells and Immune Environment in a Preclinical Model of Breast Cancer. Cancers (Basel) 2021, 13.
- 178. Gillis RD, Botteri E, Chang A, Ziegler AI, Chung NC, Pon CK, Shackleford DM, Andreassen BK, Halls ML, Baker JG, Sloan EK. Carvedilol blocks neural regulation of breast cancer progression in vivo and is associated with reduced breast cancer mortality in patients. Eur J Cancer. 2021;147:106–16.
- 179. Ganz PA, Cole SW. Expanding our therapeutic options: Beta blockers for breast cancer? J Clin Oncol. 2011;29:2612–6.
- Russo P, Del Bufalo A, Milic M, Salinaro G, Fini M, Cesario A. Cholinergic receptors as target for cancer therapy in a systems medicine perspective. Curr Mol Med. 2014;14:1126–38.
- 181. Kuol N, Davidson M, Karakkat J, Filippone RT, Veale M, Luwor R, Fraser S, Apostolopoulos V, Nurgali K: Blocking Muscarinic Receptor 3 Attenuates Tumor Growth and Decreases Immunosuppressive and Cholinergic Markers in an Orthotopic Mouse Model of Colorectal Cancer. Int J Mol Sci 2022, 24.
- Cocco E, Scaltriti M, Drilon A. NTRK fusion-positive cancers and TRK inhibitor therapy. Nat Rev Clin Oncol. 2018;15:731–47.
- Drilon A, Ou SI, Cho BC, Kim DW, Lee J, Lin JJ, Zhu VW, Ahn MJ, Camidge DR, Nguyen J, et al. Repotrectinib (TPX-0005) Is a Next-Generation ROS1/TRK/ALK Inhibitor That Potently Inhibits ROS1/TRK/ALK Solvent-Front Mutations. Cancer Discov. 2018;8:1227–36.
- 184. Fioramonti M, Fausti V, Pantano F, Iuliani M, Ribelli G, Lotti F, Pignochino Y, Grignani G, Santini D, Tonini G, Vincenzi B. Cabozantinib Affects Osteosarcoma Growth Through A Direct Effect On Tumor Cells and Modifications In Bone Microenvironment. Sci Rep. 2018;8:4177.
- 185. Italiano A, Mir O, Mathoulin-Pelissier S, Penel N, Piperno-Neumann S, Bompas E, Chevreau C, Duffaud F, Entz-Werlé N, Saada E, et al. Cabozantinib in patients with advanced Ewing sarcoma or osteosarcoma (CABONE): a multicentre, single-arm, phase 2 trial. Lancet Oncol. 2020;21:446–55.
- Takahashi K, Ehata S, Miyauchi K, Morishita Y, Miyazawa K, Miyazono K. Neurotensin receptor 1 signaling promotes pancreatic cancer progression. Mol Oncol. 2021;15:151–66.
- Li Z, You Y, Griffin N, Feng J, Shan F. Low-dose naltrexone (LDN): A promising treatment in immune-related diseases and cancer therapy. Int Immunopharmacol. 2018;61:178–84.
- Johnson DB, Manouchehri A, Haugh AM, Quach HT, Balko JM, Lebrun-Vignes B, Mammen A, Moslehi JJ, Salem JE. Neurologic toxicity associated with immune checkpoint inhibitors: a pharmacovigilance study. J Immunother Cancer. 2019;7:134.
- 189. Misawa S, Denda T, Kodama S, Suzuki T, Naito Y, Kogawa T, Takada M, Suichi T, Shiosakai K, Kuwabara S. Efficacy and safety of mirogabalin for chemotherapy-induced peripheral neuropathy: a prospective singlearm trial (MiroCIP study). BMC Cancer. 2023;23:1098.
- 190. Mravec B. Neurobiology of cancer: Definition, historical overview, and clinical implications. Cancer Med. 2022;11:903–21.
- 191. Huang Q, Hu B, Zhang P, Yuan Y, Yue S, Chen X, Liang J, Tang Z, Zhang B. Neuroscience of cancer: unraveling the complex interplay between the nervous system, the tumor and the tumor immune microenvironment. Mol Cancer. 2025;24:24.
- 192. Mravec B. Neurobiology of Cancer:Role of the Nervous System in Cancer Etiopathogenesis, Treatment, and Prevention. Springer Nature Switzerland; 2024.
- 193. Mravec B. A supportive programme for cancer patients based on knowledge of the neurobio logy of cancer. Klin Onkol. 2025;38:6–15.
- 194. Kotouček P, Enright R, Gregor Sorgerová S, Hunáková Ľ, Chlebcová V, Cholujová D, Jakubíková J, Mravec B, Naništová E, Paneková Ľ, Sedlák J. Neurobio-logy of multiple myeloma and its therapeutical use - results of the pilot study with a control arm. Klin Onkol. 2023;36:287–99.

- Xiang L, Wan H, Zhu Y. Effects of cognitive behavioral therapy on resilience among adult cancer patients: a systematic review and metaanalysis. BMC Psychiatry. 2025;25:204.
- Baydoun M, Moran C, McLennan A, Piedalue KL, Oberoi D, Carlson LE. Mindfulness-Based Interventions in Cancer Survivors: A Systematic Review of Participants' Adherence to Home Practice. Patient Prefer Adherence. 2021;15:1225–42.
- 197. Carlson LE. Mindfulness-based interventions for coping with cancer. 2016;1373:5–12.
- 198. Cahn BR, Goodman MS, Peterson CT, Maturi R, Mills PJ. Yoga, Meditation and Mind-Body Health: Increased BDNF, Cortisol Awakening Response, and Altered Inflammatory Marker Expression after a 3-Month Yoga and Meditation Retreat. Front Hum Neurosci. 2017;11:315.
- 199. Gitler A, Vanacker L, De Couck M, De Leeuw I, Gidron Y: Neuromodulation Applied to Diseases: The Case of HRV Biofeedback. J Clin Med 2022, 11.
- Leung L. Neurophysiological basis of acupuncture-induced analgesia– an updated review. J Acupunct Meridian Stud. 2012;5:261–70.
- 201. de Sousa TR, Mattos S, Marcon G, Furtado T. Duarte da Silva M: Acupuncture techniques and acupoints used in individuals under chemotherapy or radiotherapy treatment of cancer: A systematic review. J Clin Nurs. 2023;32:6917–33.
- Mao JJ, Ismaila N, Bao T, Barton D, Ben-Arye E, Garland EL, Greenlee H, Leblanc T, Lee RT, Lopez AM, et al. Integrative Medicine for Pain Management in Oncology: Society for Integrative Oncology-ASCO Guideline. J Clin Oncol. 2022;40:3998–4024.
- Epstein AS, Liou KT, Romero SAD, Baser RE, Wong G, Xiao H, Mo Z, Walker D, MacLeod J, Li Q, et al. Acupuncture vs Massage for Pain in Patients Living With Advanced Cancer: The IMPACT Randomized Clinical Trial. JAMA Netw Open. 2023;6: e2342482.
- Tian W, Zhang Y, Yu B, Jin H, Wang W, Yuan T, Yu S, Lu H. Transcutaneous electrical acupoint stimulation for alleviating pain in patients with advanced pancreatic cancer. J Cancer Res Ther. 2024;20:1334–7.
- 205. Pan C, Winkler F. Insights and opportunities at the crossroads of cancer and neuroscience. Nat Cell Biol. 2022;24:1454–60.
- Faulkner S, Jobling P, March B, Jiang CC, Hondermarck H. Tumor Neurobiology and the War of Nerves in Cancer. Cancer Discov. 2019;9:702–10.
- Engler H, Bailey MT, Engler A, Sheridan JF. Effects of repeated social stress on leukocyte distribution in bone marrow, peripheral blood and spleen. J Neuroimmunol. 2004;148:106–15.
- Engler H, Dawils L, Hoves S, Kurth S, Stevenson JR, Schauenstein K, Stefanski V. Effects of social stress on blood leukocyte distribution: the role of alpha- and beta-adrenergic mechanisms. J Neuroimmunol. 2004;156:153–62.
- 209. Murmann T, Carrillo-García C, Veit N, Courts C, Glassmann A, Janzen V, Madea B, Reinartz M, Harzen A, Nowak M, et al. Staurosporine and extracellular matrix proteins mediate the conversion of small cell lung carcinoma cells into a neuron-like phenotype. PLoS ONE. 2014;9: e86910.
- Zeng Q, Michael IP, Zhang P, Saghafinia S, Knott G, Jiao W, McCabe BD, Galván JA, Robinson HPC, Zlobec I, et al. Synaptic proximity enables NMDAR signalling to promote brain metastasis. Nature. 2019;573:526–31.

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